$\langle \rangle$	Springer Open	
-------------------	---------------	--

Search Q

Books

<u>About</u>

<u>Login</u>

Get published

**Explore** Journals

Menu 🔻

#### **Bioresources and Bioprocessing**

About Articles Submission Guidelines

#### About -

Contact **Editorial Board Editorial Board** 

#### Chairman

Jingping Qu, East China University of Science and Technology, China **Vice Chairs** Ying-Ping Zhuang, East China University of Science and Technology, China

Li-Xin Zhang, East China University of Science and Technology, China Yong-Bin Zhou, ECUST Press, China

#### **Editors-in-Chief**

Kazuyuki Shimizu, Keio University, Japan Jian-He Xu, East China University of Science and Technology, China Tingyue Gu, Ohio University, United States of America

#### **Editors**

Uttam Banerjee, National Institute of Pharmaceutical Education and Research, India Qiang Hua, East China University of Science and Technology, China Hiroyuki Imanaka, Okayama University, Japan Eun Yeol Lee, Kyung Hee University, South Korea Luo Liu, Beijing University of Chemical Technology, China Shijie Liu, SUNY College of Environmental Science and Forestry, United States of America I-Son Ng, National Cheng Kung University Yinbo Qu, Shangdong University, China Tuck Seng Wong, University of Sheffield, United Kingdom Qin Ye, East China University of Science and Technology, China Hui-Lei Yu, East China University of Science and Technology, China Lin Zhang, Tianjin University, China Yuan-Xing Zhang, East China University of Science and Technology, China Zongbao Kent Zhao, Dalian Institute of Chemical Physics, CAS, China Zhi-liang Fan, University of California, Davis, United States of America Tomohisa Hasunuma, Kobe University, Japan

#### **Editorial Board Members**

Haruyuki Atomi, Kyoto University, Japan Jin Seop Bak, Korea Advanced Institute of Science and Technology, South Korea Jie Bao, East China University of Science and Technology, China Elba P.S. Bon, Universidade Federal do Rio de Janeiro, Brazil Uwe Bornscheuer, University of Greifswald, Germany Frank Bruggeman, VU University, Netherlands Yu-Kaung Chang, Ming Chi University of Technology, China Bor-Yann Chen, National I-Lan University, China

Hong-Zhang Chen, Institute of Process Engineering, CAS, China Jian Chen, Jiangnan University, China Xin-De Chen, Guangzhou Institute of Energy Conversion, CAS, China Yijun Chen, China Pharmaceutical University, China Byung-Kwan Cho, Korea Advanced Institute of Science and Technology, South Korea Perry Chou, Waterloo University, Canada Papita Das, Jadavpur University, India Abigail Engelberth, Purdue University, United States of America Dai-Di Fan, Northwest University, China Julia Z. Fan, University of California, Davis, United States of America Xu Fang, Shandong University, China Yan Feng, Shanghai Jiaotong University, China Yingang Feng, Qingdao Institute of Biomass Energy and Bioprocess Technology, CAS, China Patrick Fickers, University of Liege, Belgium Stephen S. Fong, Virginia Commonwealth University, United States of America Marco Fraaije, University of Groningen, Netherlands Jun Ge, Tsinghua University, China Zhengrong Gu, South Dakota State University, United States of America Zheng Guo, Aarhus University, Denmark Bingfang He, Nanjing Tech University, China He Huang, Nanjing Normal University, China Zheng-Qiang Jiang, China Agricultural University, China Yong-Su Jin, University of Illinois at Urbana-Champaign, United States of America Akihiko Kondo, Kobe University, Japan Chun Li, Tsinghua University, China Yin Li, Institute of Microbiology, CAS, China Zhi Li, National University of Singapore, Singapore Wei Liao, Michigan State University, United States of America Gunnar Lidén, Lund University, Sweden Andreas Liese, Technical University of Hamburg, Germany Juan Lin, Fuzhou University, China Changsheng Liu, East China University of Science and Technology, China Liming Liu, Jiangnan University, China Tiangang Liu, Wuhan University, China You-Yan Liu, Guangxi University, China Wen-Yong Lou, South China University of Technology, China Xuefeng Lv, Qingdao Institute of Biomass Energy and Bioprocess Technology, CAS, China Fumio Matsuda, Osaka University, Japan Bo-Zhong Mu, East China University of Science and Technology, China Tamara Nazina, Research Centre of Biotechnology, Russian Academy of Sciences, Russia Takeshi Omasa, Osaka University, Japan Xuejun Pan, University of Wisconsin-Madison, United States of America Wensheng Qin, Lakehead University, Canada Ka-Yiu San, Rice University, United States of America Sang Woo Seo, Seoul National University, South Korea Haijia Su, Beijing University of Chemical Technology, China Jibin Sun, Tianjin Institute of Industrial Biotechnology, CAS, China Yajie Tang, Shandong University, China Yinjie Tang, Washington University at St. Louis, United States of America Bas Teusink, University of Amsterdam, Netherlands Johan Thevelein, KU Leuven, Belgium Dirk Tischler, Technische Universitat Bergakademie Freiberg, Germany Maobing Tu, University of Cincinnati, United States of America Nicolas Turner, The University of Manchester, United Kingdom

S. Aljoscha Wahl, Technical University of Delft, Netherlands Ping Wang, University of Minnesota, United States of America Christoph Wittmann, Saarland University, Germany Gongyuan Wei, Soochow University, China John Woodley, Technical University of Denmark, Denmark Xiaolei Wu, Peking University, China Zhilong Xiu, Dalian University of Technology, China Jianlin (Jim) Xu, Bristol-Myers Squibb Company, United States of America Sheng Yang, Institute of Plant Physiology and Ecology, CAS, China Bin Yao, Chinese Academy of Agricultural Sciences, China Hanjie Ying, Nanjing Tech University, China Hongwei Yu, Zhejiang University, China Joshua S. Yuan, Texas A&M University, United States of America Qipeng Yuan, Beijing University of Chemical Technology, China Yingjin Yuan, Tianjin University, China An-Ping Zeng, Hamburg University of Technology, Germany Huimin Zhao, University of Illinois at Urbana-Champaign, United States of America Liming Zhao, East China University of Science and Technology, China Lishan Zhao, Amyris, Inc., United States of America Xueli Zhang, Tianjin Institute of Industrial Biotechnology, CAS, China Wei Zhang, Flinders University, Australia Yuguo Zheng, Zhejiang University of Technology, China Jian-Jiang Zhong, Shanghai Jiaotong University, China Jiahai Zhou, Shenzhen Institutes of Advanced Technology, CAS, China Kang Zhou, National University of Singapore, Singapore Yang Zhu, Wageningen University & Research, Netherlands

#### **Young Members**

Yunpeng Bai, East China University of Science and Technology, China Menghao Cai, East China University of Science and Technology, China Bigiang Chen, Beijing University of Chemical Technology, China Kequan Chen, Nanjing Tech University, China Yongzheng Chen, Zunyi Medical University, China Zhen Chen, Tsinghua University, China Changzheng Cui, East China University of Science and Technology, China Jian-Hua Fan, East China University of Science and Technology, China Hao Fang, Northwest A&F University, China Qiang Fei, Xi'an Jiaotong University, China Kai Guo, Nanjing Tech University, China Yu-Cai He, Changzhou University, China Shih-Hsin Ho, Harbin Institute of Technology, China Roman Holic, Slovak Academy of Sciences, Slovak Haitong Hou, University of Colorado, United States of America Jinguang Hu, University of Calgary, Canada Xiao-Jun Ji, Nanjing Tech University, China Mingjie Jin, Nanjing University of Science and Technology, China Donghyuk Kim, Ulsan National Institute of Science and Technology, South Korea Xu-Dong Kong, Shanghai Jiaotong University, China Ai-Tao Li, Hubei University, China Bing-Zhi Li, Tianjin University, China Ning Li, South China University of Technology, China Quan-Shun Li, Jilin University, China Shengying Li, Shandong University, China Pan Liao, Purdue University, United States of America

Sen Lin, East China University of Science and Technology, China Shuangjun Lin, Shanghai Jiaotong University, China Yuan Lu, Tsinghua University, China Yunzi Luo, Tianjin University, China Yongqing Lv, Beijing University of Chemical Technology, China Ye Ni, Jiangnan University, China Yao Nie, Jiangnan University, China Xudong Qu, Shanghai Jiaotong University, China Shu Quan, East China University of Science and Technology, China Jinyou Shen, Nanjing University of Science and Technology, China Li Shuai, Fujian A&F University, China Hao Song, Tianjin University, China Qian Sui, East China University of Science and Technology, China Zhou-Tong Sun, Tianjin Institute of Industrial Biotechnology, CAS, China Hongzhi Tang, Shanghai Jiaotong University, China Dan Wang, Chongqing University, China Jianbo Wang, Hunan Normal University, China Shi-An Wang, Qingdao Institute of Biomass Energy and Bioprocess Technology, CAS, China Wei Wang, East China University of Science and Technology, China Yajun Wang, Zhejiang University of Technology, China Yong Wang, Institute of Plant Physiology and Ecology, CAS, China Bian Wu, Institute of Microbiology, CAS, China Hui Wu, East China University of Science and Technology, China Jianping Wu, Zhejiang University, China Qi Wu, Zhejiang University, China Shuke Wu, University of Greifswald, Germany Zhongliu Wu, Chengdu Institute of Biology, CAS, China Jingli Xie, East China University of Science and Technology, China Peng Xu, University of Maryland, Baltimore County, United States of America Chuang Xue, Dalian University of Technology, China Yang-Chun Yong, Jiangsu University, China Chun You, Tianjin Institute of Industrial Biotechnology, CAS, China Xianhai Zeng, Xiamen University, China Cheng Zhang, KTH-Royal Institute of Technology, Sweden Haoran Zhang, Rutgers University, United States of America Jian-Dong Zhang, Taiyuan University of Technology, China Xuebing Zhao, Tsinghua University, China Yuzheng Zhao, East China University of Science and Technology, China Gao-Wei Zheng, East China University of Science and Technology, China Renchao Zheng, Zhejiang University of Technology, China Yongjin Zhou, Dalian Institute of Chemical Physics, CAS, China Leilei Zhu, Tianjin Institute of Industrial Biotechnology, CAS, China Ning Zhu, Nanjing Tech University, China

#### **Advisory Members**

Yasuhisa Asano, Toyama Prefectural University, Japan Virendra Bisaria, Indian Institute of Technology Delhi, India Jo-Shu Chang, National Cheng Kung University, China Muhammed Umar Dahot, University of Sindh, Pakistan Zixin Deng, Shanghai Jiaotong University, China Wei-Shou Hu, University of Minnesota, United States of America Tadayuki Imanaka, Ritsumeikan University, Japan Romas Kazlauskas, University of Minnesota, United States of America Byung-Gee Kim, Seoul National University, South Korea

Dehua Liu, Tsinghua University, China Xiaojun Ma, Dalian Institute of Chemical Physics, CAS, China Ashok Mulchandani, University of California, Riverside, United States of America Teruyuki Nagamune, The University of Tokyo, Japan Pingkai Ouyang, Nanjing Tech University, China Sunghoon Park, Pusan National University, South Korea Manfred Reetz, Max-Planck-Institut für Kohlenforschung, Germany Peer Schenk, The University of Queensland, Australia Gregory Stephanopoulos, Massachusetts Institute of Technology, United States of America Runcang Sun, Beijing Forestry University, China Vytas Svedas, Lomonosov Moscow State University, Russia Tianwei Tan, Beijing University of Chemical Technology, China Shan-Tung Tu, East China University of Science and Technology, China Jin Chuan Wu, Institute of Chemical & Engineering Sciences, Singapore Yan Xu, Jiangnan University, China Jiwon Yang, Korea Advanced Institute of Science and Technology, South Korea Lirong Yang, Zhejiang University, China Shang-Tian Yang, Ohio State University, United States of America Kaiming Ye, Binghamton University, United States of America Hanging Yu, University of Science and Technology of China, China Percival Zhang, Tianjin Institute of Industrial Biotechnology, CAS, China

Submit manuscript

#### Editorial Board

Sign up for article alerts and news from this journal

Affiliated with



*Bioresources and Bioprocessing* is associated with the <u>State Key Laboratory of Bioreactor Engineering</u>, East China University of Science and Technology.

### Annual Journal Metrics

#### 2022 Citation Impact

4.6 - 2-year Impact Factor6.1 - 5-year Impact Factor1.232 - SNIP (Source Normalized Impact per Paper)0.766 - SJR (SCImago Journal Rank)

#### 2022 Speed

6 days submission to first editorial decision for all manuscripts (Median) 80 days submission to accept (Median)

#### 2022 Usage

897,118 downloads 131 Altmetric mentions ISSN: 2197-4365 (electronic)

## This journal is indexed by

- SCOPUS
- Science Citation Index Expanded
- Google Scholar
- CNKI
- DOAJ
- EBSCO Discovery Service
- EBSCO TOC Premier
- OCLC WorldCat Discovery Service
- ProQuest Biological Science Database
- ProQuest Materials Science & Engineering Database
- ProQuest Natural Science Collection
- ProQuest SciTech Premium Collection
- ProQuest Technology Collection
- ProQuest-ExLibris Primo
- ProQuest-ExLibris Summon

Support and Contact	Terms and conditions			
Jobs	Privacy statement			
Language editing for authors	<u>Accessibility</u>			
Scientific editing for authors	<u>Cookies</u>			
Leave feedback				

Follow SpringerOpen

**(f**)

By using this website, you agree to our <u>Terms and Conditions</u>, <u>Your US state privacy rights</u>, <u>Privacy statement</u> and <u>Cookies</u> policy. <u>Your privacy choices/Manage cookies</u> we use in the preference centre.

#### **SPRINGER NATURE**

© 2023 BioMed Central Ltd unless otherwise stated. Part of Springer Nature.

10/27/23, 12:17 PM		Bioresources and Bioprocessing   Articles					
Der Springer Open		Search $Q$	<u>Get published</u> <sup>Menu</sup> ▼	Explore Journals	<u>Books</u>	<u>About</u>	<u>Login</u>
Bioresources and B	ioprocessing						
About <u>Articles</u> Submissic	n Guidelines						
Articles  Collections Reviewer Acknowledger Articles	<u>nents</u>						
Search by keyword	Search by citation						
Search Bioresources and Bioprocessing						Volume 9 (	2022) 🗸
Search <b>131</b> result(s)							
within Volume 9	of Bioresource	es and Bioproces	sing				
Page 1 of 3					So	Newes	st first ∨

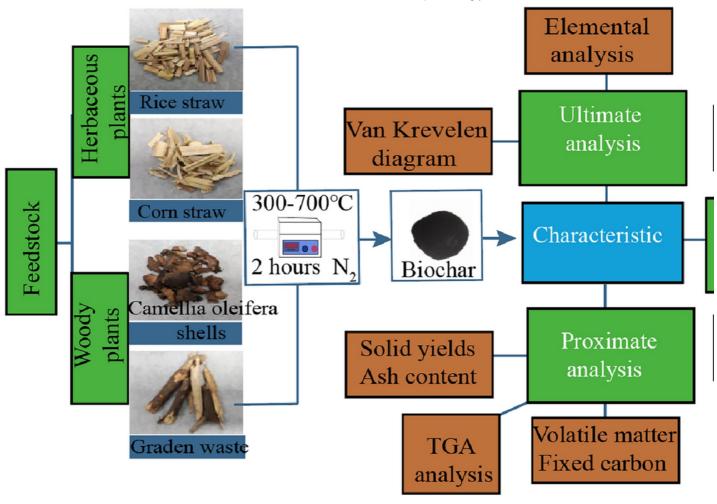
<u>Influence of pyrolysis temperature on the physicochemical properties of biochars obtained from herbaceous and woody plants</u>

This work aimed to investigate the effect of pyrolysis temperature on the yield and properties of biochars synthesized from herbaceous and woody plants. Four typical materials, including two herbaceous plants ...

Panfeng Tu, Guanlin Zhang, Guoqiang Wei, Juan Li, Yongquan Li, Lifang Deng and Haoran Yuan

Bioresources and Bioprocessing 2022 9:131

Research Published on: 22 December 2022



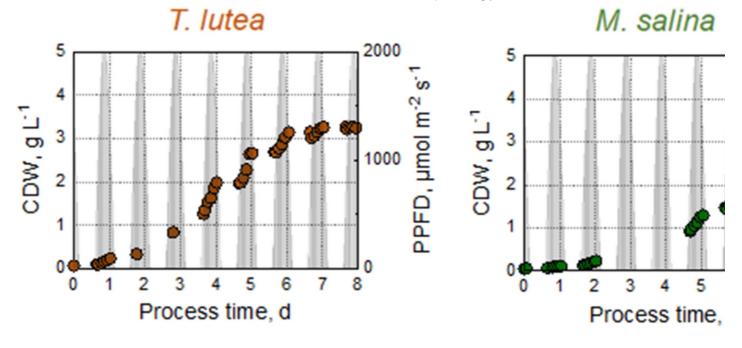
## <u>Simultaneous photoautotrophic production of DHA and EPA by *Tisochrysis lutea* and *Microchloropsis salina* in co-culture</u>

Marine microalgae have received much attention as a sustainable source of the two health beneficial omega-3-fatty acids docosahexaenoic acid (DHA, C22:6) and eicosapentaenoic acid (EPA, C20:5). However, photoa...

Anna-Lena Thurn, Anna Stock, Sebastian Gerwald and Dirk Weuster-Botz

Bioresources and Bioprocessing 2022 9:130

Research Published on: 19 December 2022



### Dissecting the essential role of N-glycosylation in catalytic performance of xanthan lyase

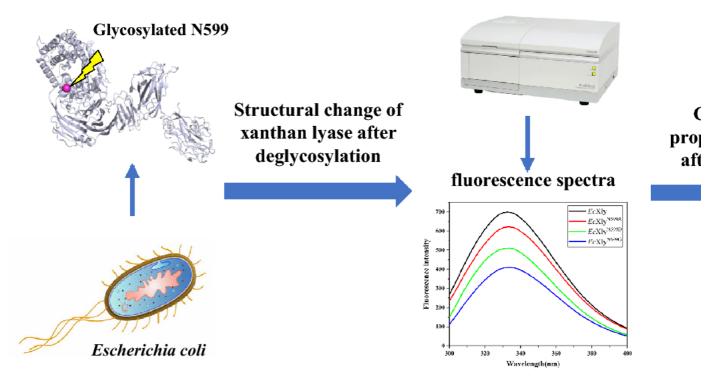
Modified xanthan produced by xanthan lyase has broad application prospects in the food industry. However, the catalytic performance of xanthan lyase still needs to be improved through rational design. To addre...

Jingjing Zhao, Qian Wang, Xin Ni, Shaonian Shen, Chenchen Nan, Xianzhen Li, Xiaoyi Chen and Fan Yang

Bioresources and Bioprocessing 2022 9:129

Research Published on: 16 December 2022

> Full Text > PDF



Tertiary structure change caused by glycosylation

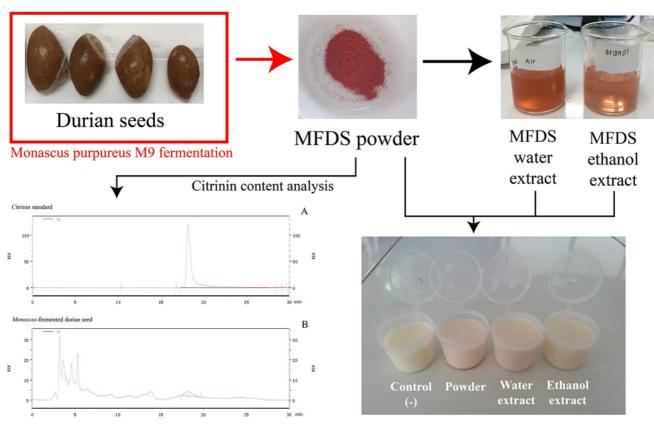
## <u>Utilization of durian seed for *Monascus* fermentation and its application as a functional ingredient</u> <u>in yogurt</u>

As a widely consumed fermented milk product, yogurt undergoes constant development to increase its functional properties. *Monascus purpureus*-fermented durian seed, which has been proven to possess antioxidative p...

Ignatius Srianta, Indah Kuswardani, Susana Ristiarini, Netty Kusumawati, Laura Godelive and Ira Nugerahani

Bioresources and Bioprocessing 2022 9:128

Research Published on: 14 December 2022



# <u>Protective effects of *Lactobacillus reuteri* SJ-47 strain exopolysaccharides on human skin fibroblasts damaged by UVA radiation</u>

Ultraviolet rays in sunlight can cause skin damage and premature aging. This study demonstrates that *Lactobacillus reuteri* SJ-47 strain exopolysaccharides (EPS) protect human skin fibroblasts (HSF) under UVA radi...

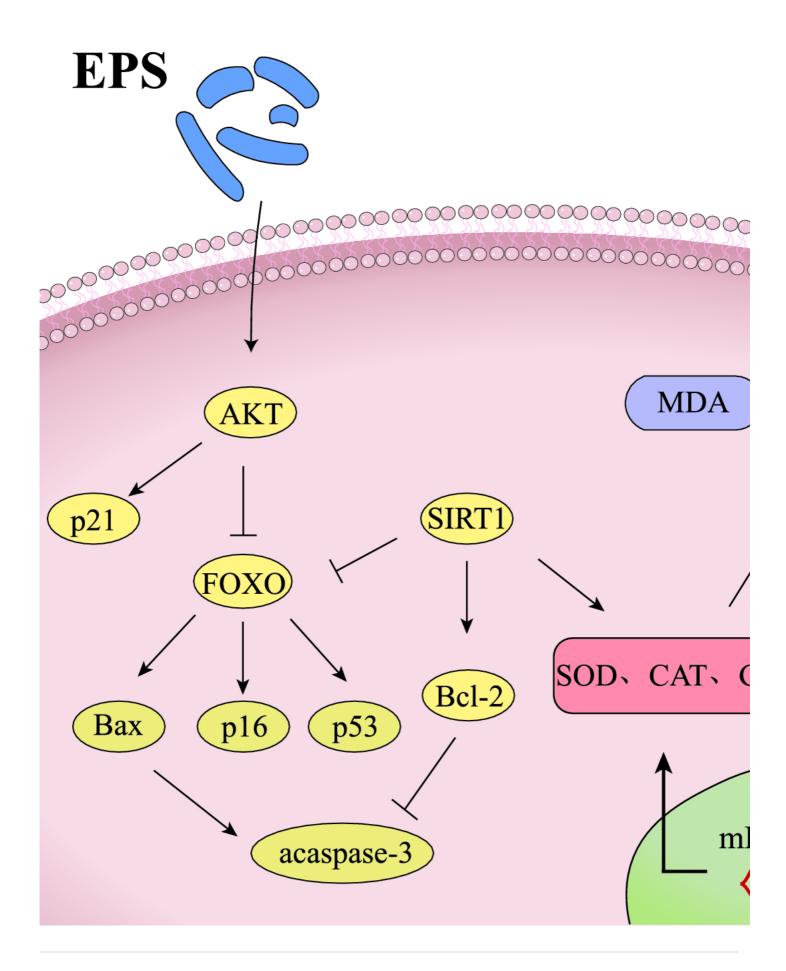
Jingsha Zhao, Hao Fu, Yongtao Zhang, Meng Li, Dongdong Wang, Dan Zhao, Jiachan Zhang and Changtao Wang

Bioresources and Bioprocessing 2022 9:127

Research Published on: 14 December 2022

> Full Text > PDF

Consis



Transcription regulation strategies in methylotrophs: progress and challenges

#### 10/27/23, 12:17 PM

#### Bioresources and Bioprocessing | Articles

As a promising industrial microorganism, methylotroph is capable of using methane or methanol as the sole carbon source natively, which has been utilized in the biosynthesis of various bioproducts. However, th...

Xiaohan Huang, Qiaoqiao Song, Shuqi Guo and Qiang Fei

Bioresources and Bioprocessing 2022 9:126

Review Published on: 12 December 2022

> Full Text > PDF

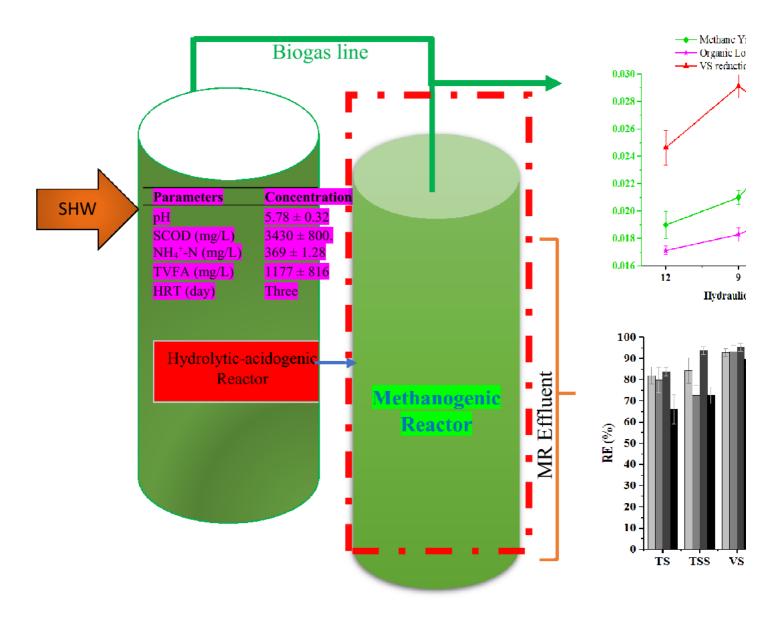
### <u>Optimization of operating parameters for biogas production using two-phase bench-scale</u> <u>anaerobic digestion of slaughterhouse wastewater: Focus on methanogenic step</u>

The objective of the present study was an optimization of operating parameters and the performance of the methanogenesis reactor in phased anaerobic digestion (AD) of slaughterhouse wastewater at 37.5°C. Accor...

Dejene Tsegaye and Seyoum Leta

Bioresources and Bioprocessing 2022 9:125

Research Published on: 9 December 2022



## <u>Synthetic biology promotes the capture of CO2 to produce fatty acid derivatives in microbial cell</u> factories

Environmental problems such as greenhouse effect, the consumption of fossil energy, and the increase of human demand for energy are becoming more and more serious, which force researcher to turn their attentio...

Xiaofang Liu, Hangyu Luo, Dayong Yu, Jinyu Tan, Junfa Yuan and Hu Li

Bioresources and Bioprocessing 2022 9:124

Review Published on: 5 December 2022

> Full Text > PDF

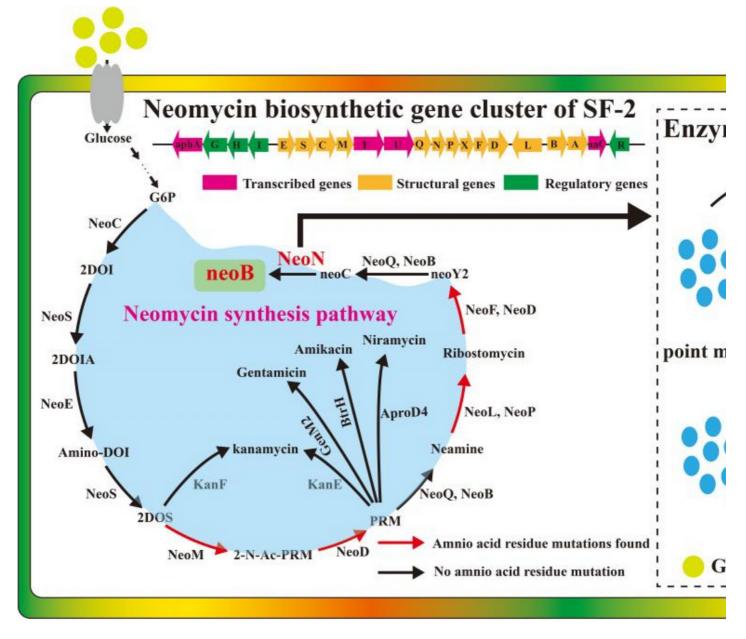
## <u>Point mutation of V252 in neomycin C epimerase enlarges substrate-binding pocket and improves</u> <u>neomycin B accumulation in *Streptomyces fradiae*</u>

Neomycin, an aminoglycoside antibiotic with broad-spectrum antibacterial resistance, is widely used in pharmaceutical and agricultural fields. However, separation and purification of neomycin B as an active su...

Xiangfei Li, Fei Yu, Fang Wang, Sang Wang, Rumeng Han, Yihan Cheng, Ming Zhao, Junfeng Sun and Zhenglian Xue

Bioresources and Bioprocessing 2022 9:123

Research Published on: 5 December 2022



## <u>An overview of torrefied bioresource briquettes: quality-influencing parameters, enhancement</u> <u>through torrefaction and applications</u>

In recent years, the need for clean, viable and sustainable source of alternative fuel is on the rampage in the global space due to the challenges posed by human factors including fossil induced emissions, fue...

M. A. Waheed, O. A. Akogun and C. C. Enweremadu

Bioresources and Bioprocessing 2022 9:122

Review Published on: 28 November 2022

> Full Text > PDF

# Efficient 1,3-dihydroxyacetone biosynthesis in *Gluconobacter oxydans* using metabolic engineering and a fed-batch strategy

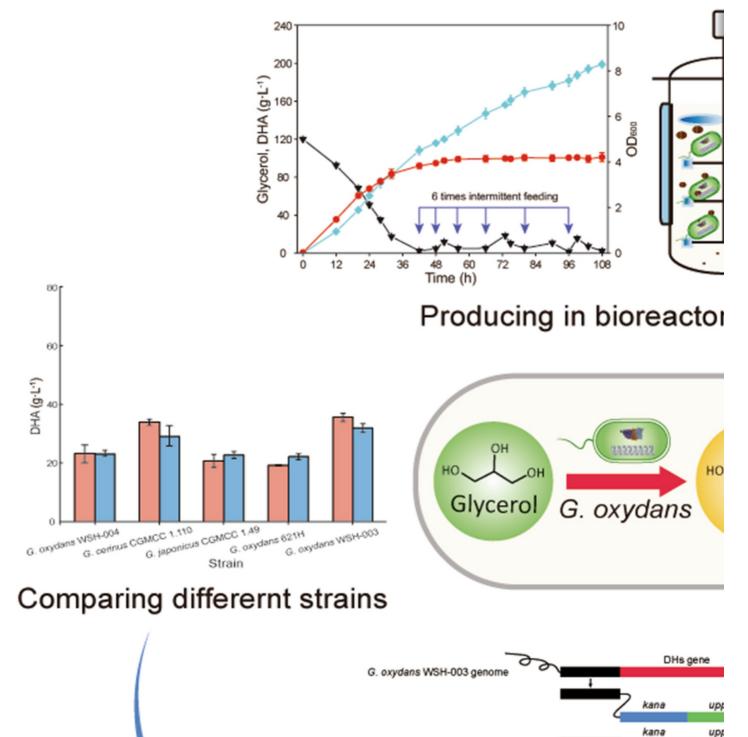
1,3-Dihydroxyacetone (DHA) is a commercially important chemical and widely used in cosmetics, pharmaceuticals, and food industries as it prevents excessive water evaporation, and provides anti-ultraviolet radi...

#### 10/27/23, 12:17 PM

Weizhu Zeng, Xiaoyu Shan, Li Liu and Jingwen Zhou

Bioresources and Bioprocessing 2022 9:121

Research | Published on: 26 November 2022



## Knocking out dehydrogen

<u>Enhancing menaquinone-7 biosynthesis by adaptive evolution of *Bacillus natto* through chemical modulator</u>

G. oxydans WSH-003 genor

G. oxydans WSH-003 genome

G. oxydans WSH-003 genome

DHs gene

kana

#### 10/27/23, 12:17 PM

#### Bioresources and Bioprocessing | Articles

Menaquinone-7 (MK-7) is a kind of vitamin K2 playing an important role in the treatment and prevention of cardiovascular disease, osteoporosis and arterial calcification. The purpose of this study is to establ...

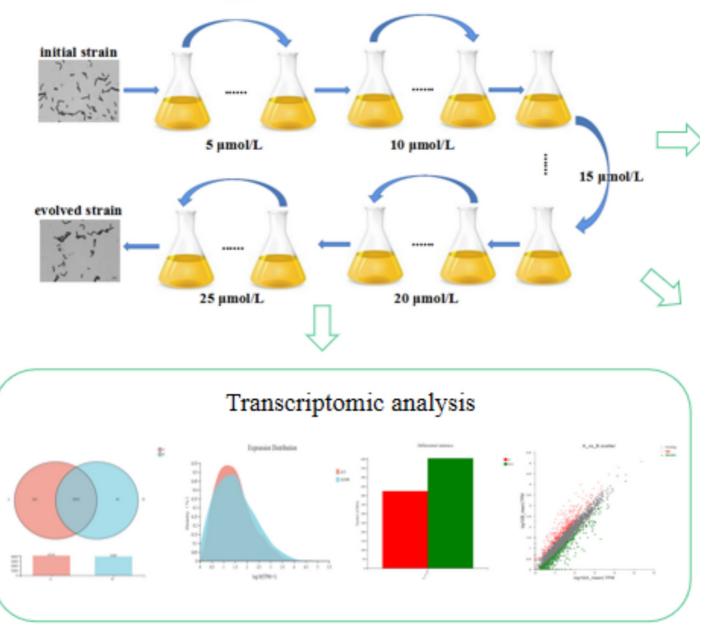
Bei Zhang, Cheng Peng, Jianyao Lu, Xuechao Hu and Lujing Ren

Bioresources and Bioprocessing 2022 9:120

Research | Published on: 22 November 2022

### > Full Text > PDF

## Adaptive evolution on glyphosate



## <u>Expression, characterization, and application potentiality evaluation of recombinant human-like</u> <u>collagen in *Pichia pastoris*</u>

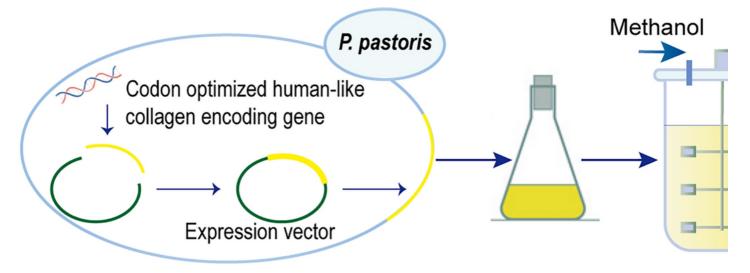
Collagen is a biofunctional protein that has been widely used in many fields, including but not limited to biomedical, cosmetics and skin care, food, and novel materials. Recombinant collagen has great potenti...

Lingling Ma, Xiaolin Liang, Shiqin Yu and Jingwen Zhou

Bioresources and Bioprocessing 2022 9:119

Research Published on: 17 November 2022

#### > Full Text > PDF





Collagen sponge

## Minos and Restless transposon insertion mutagenesis of psychrotrophic fungus for red pigment synthesis adaptive to normal temperature

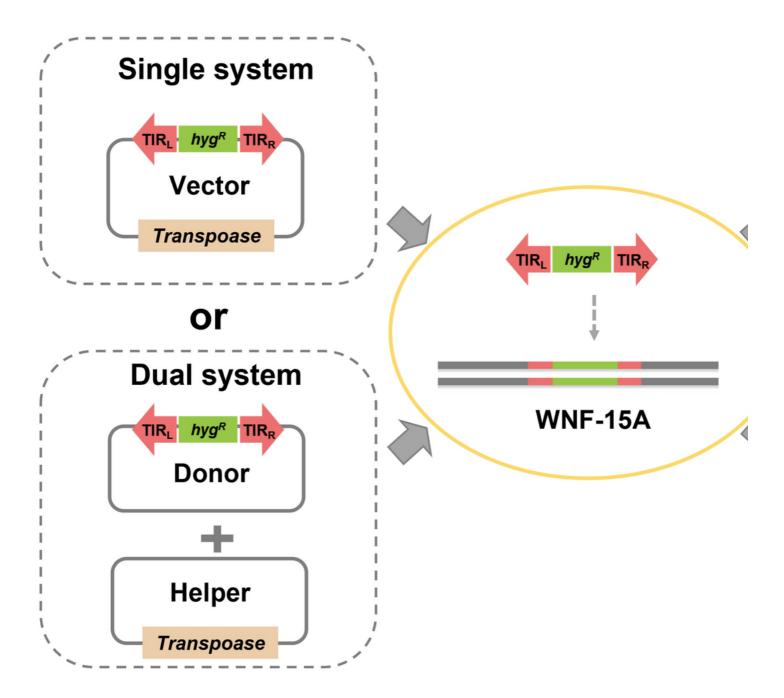
The polar psychrotrophic fungus Geomyces sp. WNF-15A can produce high-quality natural red pigment for the potential use as edible pigment. However, it shows low-temperature-dependent synthesis of red pigment, whi...

Fengning Lu, Yanna Ren, Lulu Ding, Jian Lu, Xiangshan Zhou, Haifeng Liu, Nengfei Wang and Menghao Cai

Bioresources and Bioprocessing 2022 9:118

Research Published on: 4 November 2022

- 10:6 The <u>Correction to this article</u> has been published in *Bioresources and Bioprocessing* 2023
- > Full Text > PDF



## Plasmids Construction

## **Random Integration**

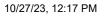
## <u>High-efficiency secretory expression and characterization of the recombinant type III human-like</u> <u>collagen in *Pichia pastoris*</u>

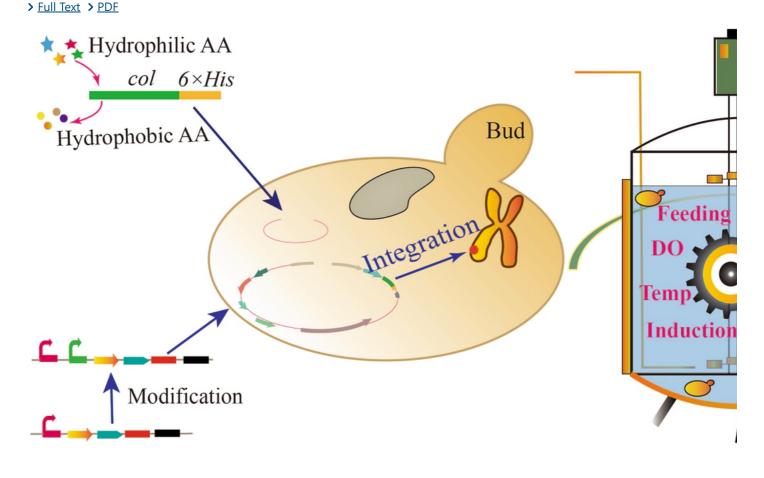
Collagen, the highest content protein in the body, has irreplaceable biological functions, and it is widespread concerned in food, beauty, and medicine with great market demand. The gene encoding the recombina...

Zhi-Xiang Xiang, Jin-Song Gong, Jin-Hao Shi, Chun-Fang Liu, Heng Li, Chang Su, Min Jiang, Zheng-Hong Xu and Jin-Song Shi

Bioresources and Bioprocessing 2022 9:117

Research Published on: 4 November 2022





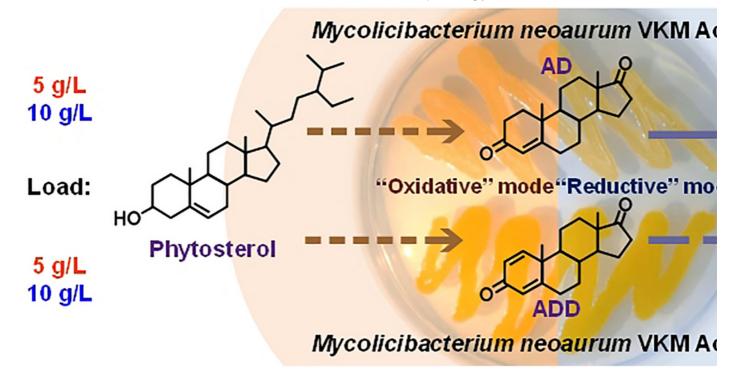
## <u>Bioproduction of testosterone from phytosterol by *Mycolicibacterium neoaurum* strains: "onepot", two modes</u>

The main male hormone, testosterone is obtained from cheap and readily available phytosterol using the strains of *Mycolicibacterium neoaurum* VKM Ac-1815D, or Ac-1816D. During the first "oxidative" stage, phytoste...

Daria N. Tekucheva, Vera M. Nikolayeva, Mikhail V. Karpov, Tatiana A. Timakova, Andrey V. Shutov and Marina V. Donova

Bioresources and Bioprocessing 2022 9:116

Research Published on: 4 November 2022



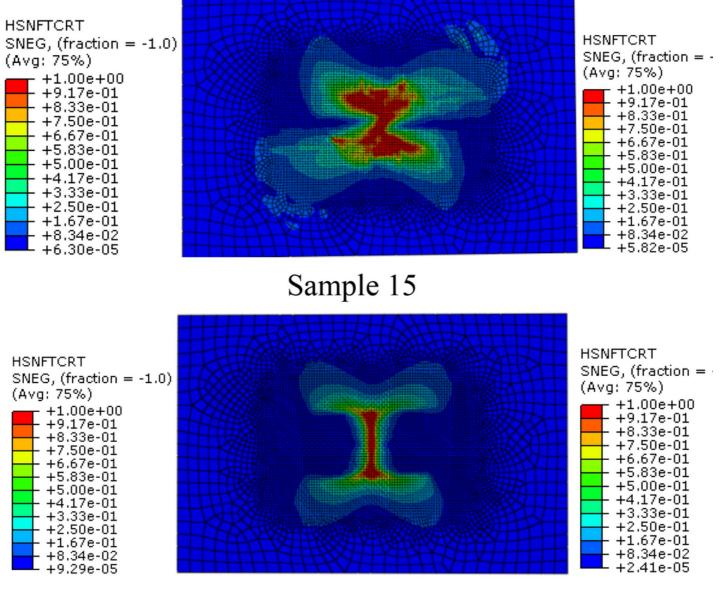
## <u>Bionic design based on micro-nano structure of osteon and its low-velocity impact damage</u> <u>behavior</u>

It is found that the osteon is composed of thin and thick lamellae which are periodic and approximately concentric, every 5 lamellae is a cycle, the periodic helix angle of mineralized collagen fibers in two a...

Yuxi Liu, Aihua Li, Yanhua Li and Song Chen

Bioresources and Bioprocessing 2022 9:115

Research Published on: 2 November 2022



Sample 60

## <u>Chemical composition and bioactivity of oilseed cake extracts obtained by subcritical and modified</u> <u>subcritical water</u>

Recovery of bioactive compounds from biowaste is gaining more and more interest in circular economy models. The oilseed cakes are usually insufficiently exploited by most technologies since they represent valu...

Jaroslava Švarc-Gajić, Francisca Rodrigues, Manuela M. Moreira, Cristina Delerue-Matos, Simone Morais, Olena Dorosh, Ana Margarida Silva, Andrea Bassani, Valentin Dzedik and Giorgia Spigno

Bioresources and Bioprocessing 2022 9:114

Research Published on: 29 October 2022



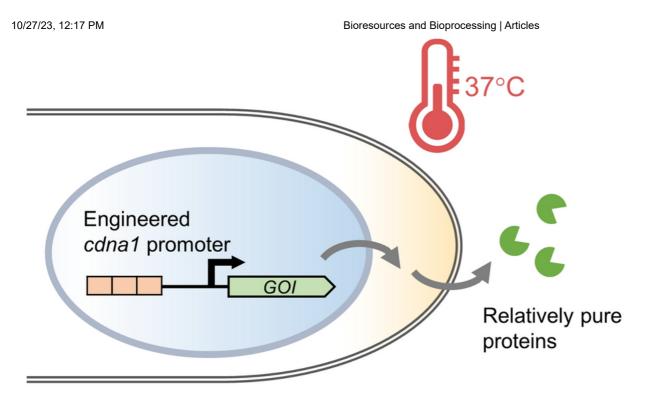
## <u>Genetic engineering and raising temperature enhance recombinant protein production with the</u> <u>cdna1 promoter in Trichoderma reesei</u>

The fungus *Trichoderma reesei* is a powerful host for secreted production of proteins. The promoter of *cdna1* gene, which encodes a small basic protein of unknown function and high expression, is commonly used for ...

Shanshan Jiang, Yue Wang, Qin Liu, Qinqin Zhao, Liwei Gao, Xin Song, Xuezhi Li, Yinbo Qu and Guodong Liu

Bioresources and Bioprocessing 2022 9:113

Research Published on: 29 October 2022



Trichoderma reesei

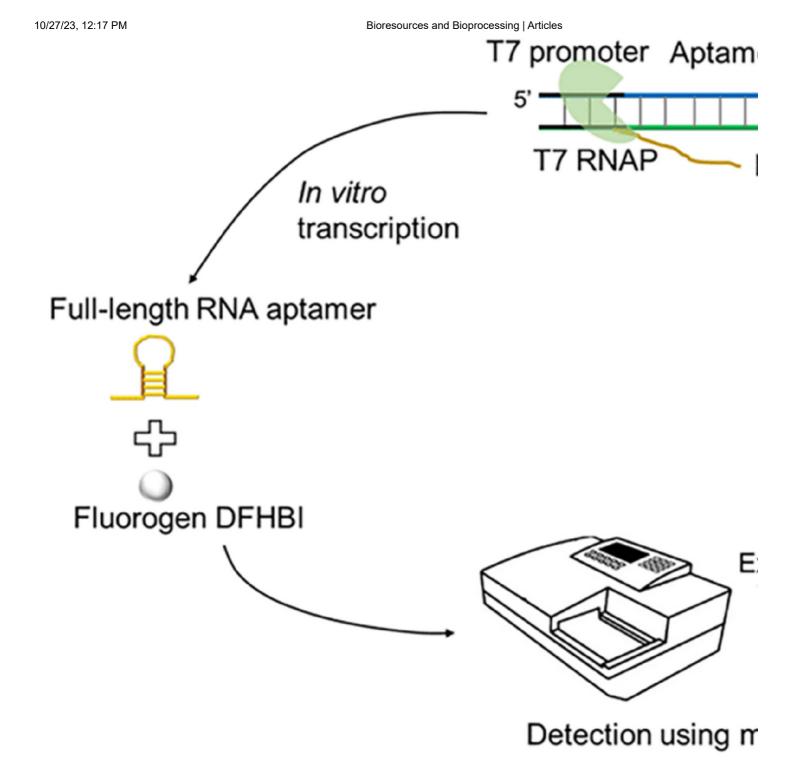
## <u>High-throughput iSpinach fluorescent aptamer-based real-time monitoring of in vitro</u> <u>transcription</u>

In vitro transcription (IVT) is an essential technique for RNA synthesis. Methods for the accurate and rapid screening of IVT conditions will facilitate RNA polymerase engineering, promoter optimization, and s...

Weitong Qin, Liang Li, Fan Yang, Siyuan Wang and Guang-Yu Yang

Bioresources and Bioprocessing 2022 9:112

Research Published on: 27 October 2022



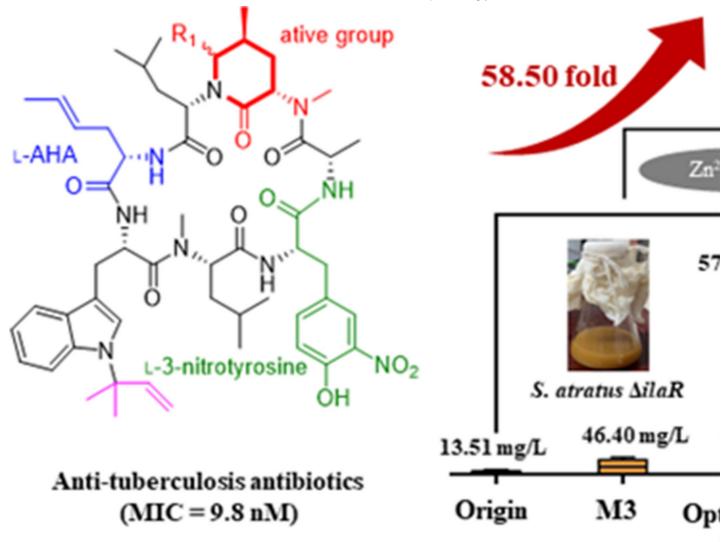
## <u>Combinatorial strategies for production improvement of anti-tuberculosis antibiotics ilamycins</u> <u> $E_1/E_2$ from deep sea-derived Streptomyces atratus SCSIO ZH16 $\Delta ilaR$ </u>

Ilamycins E<sub>1</sub>/E<sub>2</sub> are novel cyclic heptapeptides from *Streptomyces atratus* SCSIO ZH16, which have the MIC value of 9.8 nM against *Mycobacterium tuberculosis* H37Rv. However, the lower fermentative titer of ilamycins...

Yunfei Zhu, Gaofan Zheng, Xiujuan Xin, Junying Ma, Jianhua Ju and Faliang An

Bioresources and Bioprocessing 2022 9:111

Research Published on: 22 October 2022



## <u>Dynamic changes in community structure and degradation performance of a bacterial consortium</u> <u>MMBC-1 during the subculturing revival reveal the potential decomposers of lignocellulose</u>

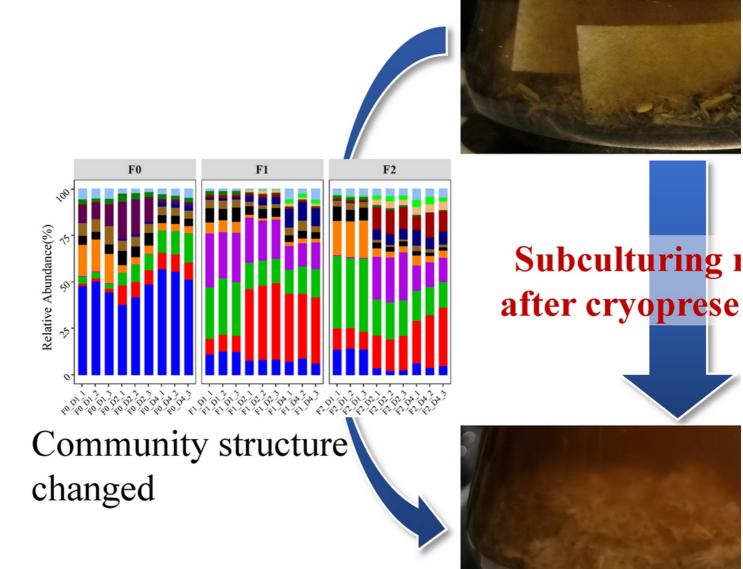
Bacterial consortium is an important source of lignocellulolytic strains, but it is still a challenge to distinguish the direct decomposers of lignocellulose from other bacteria in such a complex community. Th...

Jingrong Zhu, Jiawen Liu, Weilin Li, Yunrui Ru, Di Sun, Cong Liu, Zongyun Li and Weijie Liu

Bioresources and Bioprocessing 2022 9:110

Research Published on: 22 October 2022

# Lignocellulolytic c



# Identifying potential decomposers of lign

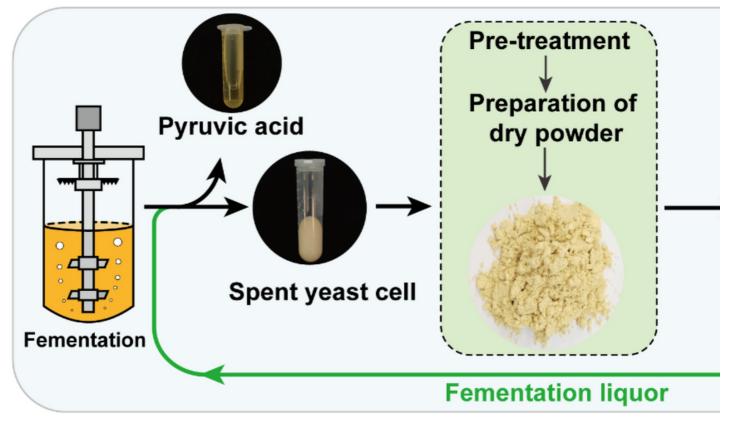
# <u>Production of pyruvic acid with *Candida glabrata* using self-fermenting spent yeast cell dry powder as a seed nitrogen source</u>

Pyruvic acid is an important organic acid and a key industrial raw material. It is widely used in the chemical, agricultural, and food fields. *Candida glabrata* is the preferred strain for pyruvic acid production....

Qiyuan Lu, Xiaoyu Shan, Weizhu Zeng and Jingwen Zhou

Bioresources and Bioprocessing 2022 9:109

Research Published on: 17 October 2022



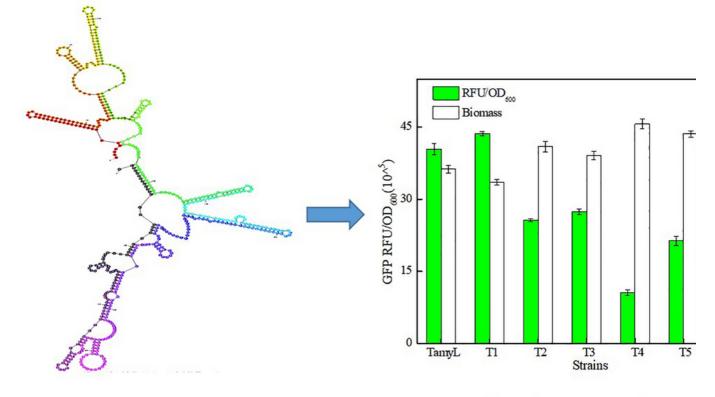
## <u>Minimization and optimization of α-amylase terminator for heterologous protein production in</u> <u>Bacillus licheniformis</u>

Terminators serve as the regulatory role in gene transcription termination; however, few researches about terminator optimization have been conducted, which leads to the lack of available and universal termina...

Yi Rao, Jingyao Yang, Jiaqi Wang, Xinyuan Yang, Mengxi Zhang, Yangyang Zhan, Xin Ma, Dongbo Cai, Zhangqian Wang and Shouwen Chen

Bioresources and Bioprocessing 2022 9:108

Research Published on: 10 October 2022



## Secondary structure of Terminator

## **Terminator screening**

### A novel sustainable platform for scaled manufacturing of double-stranded RNA biopesticides

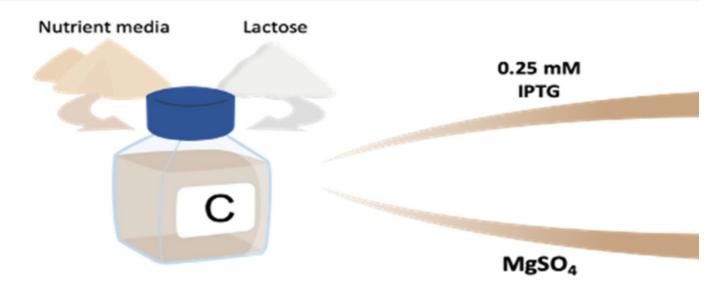
RNA interference (RNAi) represents one of the most conserved pathways evolved by eukaryotic cells for regulating gene expression. RNAi utilises non-translatable double-stranded RNA (dsRNA) molecules to sequest...

Alison Obinna Nwokeoji, Eleojo Ahuva Nwokeoji, Tachung Chou and Abou Togola

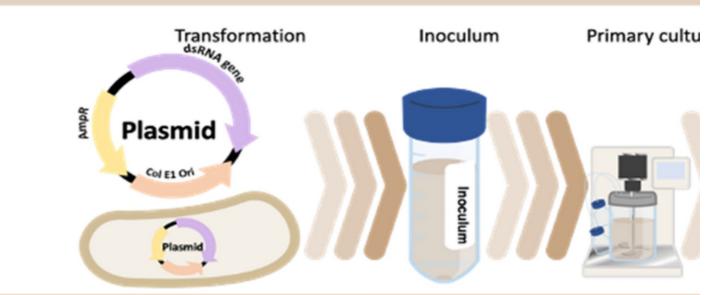
Bioresources and Bioprocessing 2022 9:107

Research Published on: 6 October 2022

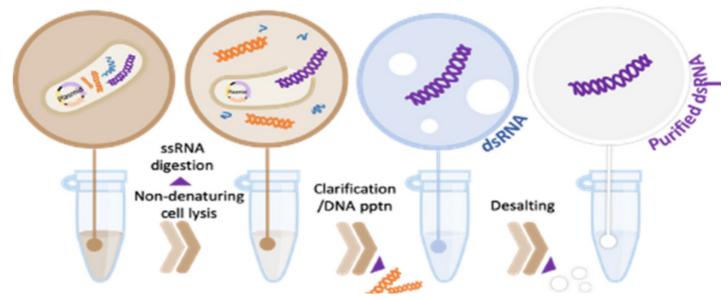
## **Autoinduction media formulation**



**Microbial fermentation** 



dsRNA purification





Nucleotides /salts

## <u>RETRACTED ARTICLE: Mutagenesis combined with fermentation optimization to enhance</u> <u>gibberellic acid GA3 yield in *Fusarium fujikuroi*</u>

Gibberellic acid (GA3) is a plant growth hormone that plays an important role in the production of crops, fruits, and vegetables with a wide market share. Due to intrinsic advantages, liquid fermentation of *Fusar*...

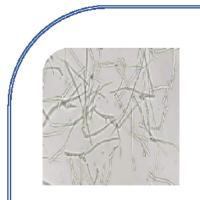
Ya-Wen Li, Cai-Ling Yang, Hui Peng, Zhi-Kui Nie, Tian-Qiong Shi and He Huang

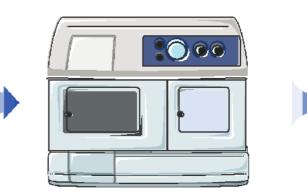
F. fujikuroi NJtech 02

Bioresources and Bioprocessing 2022 9:106

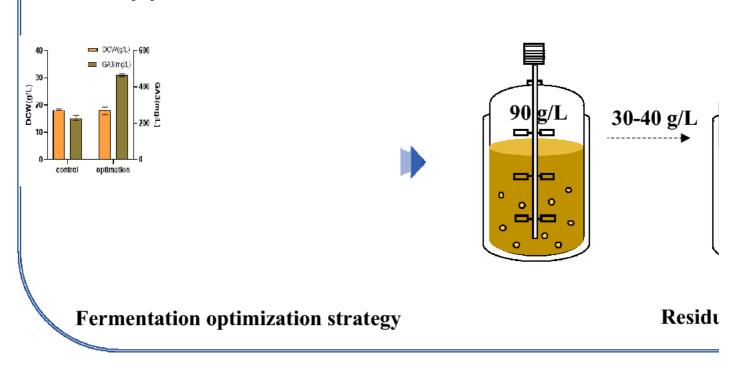
Research Published on: 5 October 2022

#### > Full Text > PDF





## **ARTP** mutation



## <u>Production of xylose through enzymatic hydrolysis of glucuronoarabinoxylan from brewers' spent</u> <u>grain</u>

#### 10/27/23, 12:17 PM

#### Bioresources and Bioprocessing | Articles

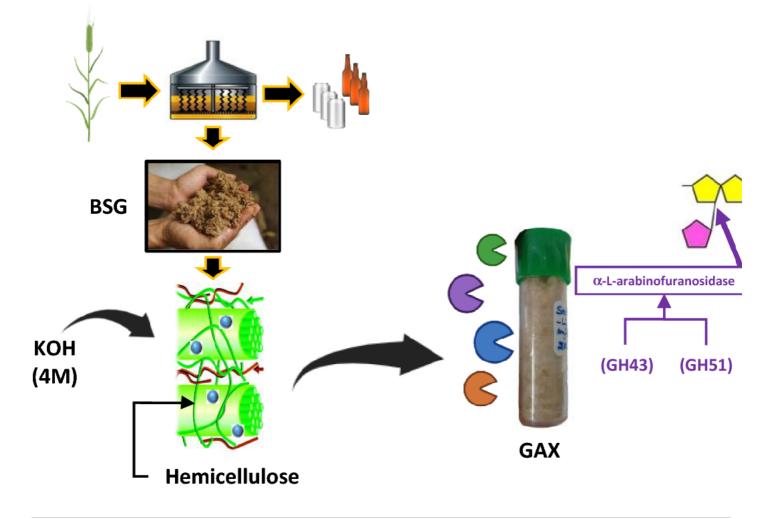
Xylose is an abundant bioresource for obtaining diverse chemicals and added-value products. The production of xylose from green alternatives like enzymatic hydrolysis is an important step in a biorefinery cont...

Lilia C. Rojas-Pérez, Paulo C. Narváez-Rincón, M. Angélica M. Rocha, Elisabete Coelho and Manuel A. Coimbra

Bioresources and Bioprocessing 2022 9:105

Research Published on: 4 October 2022

#### > Full Text > PDF



## <u>Transcriptomic and proteomic analyses provide insights into the adaptive responses to the</u> <u>combined impact of salinity and alkalinity in *Gymnocypris przewalskii*</u>

*Gymnocypris przewalskii* is the only high-land endemic teleost living in Qinghai Lake, the largest saline–alkaline lake in China. Its osmoregulatory physiology remains elusive due to a lack of precise identificati...

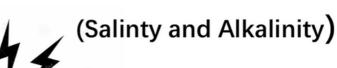
Fulei Wei, Jian Liang, Wengen Tian, Luxian Yu, Zhaohui Feng and Qiang Hua

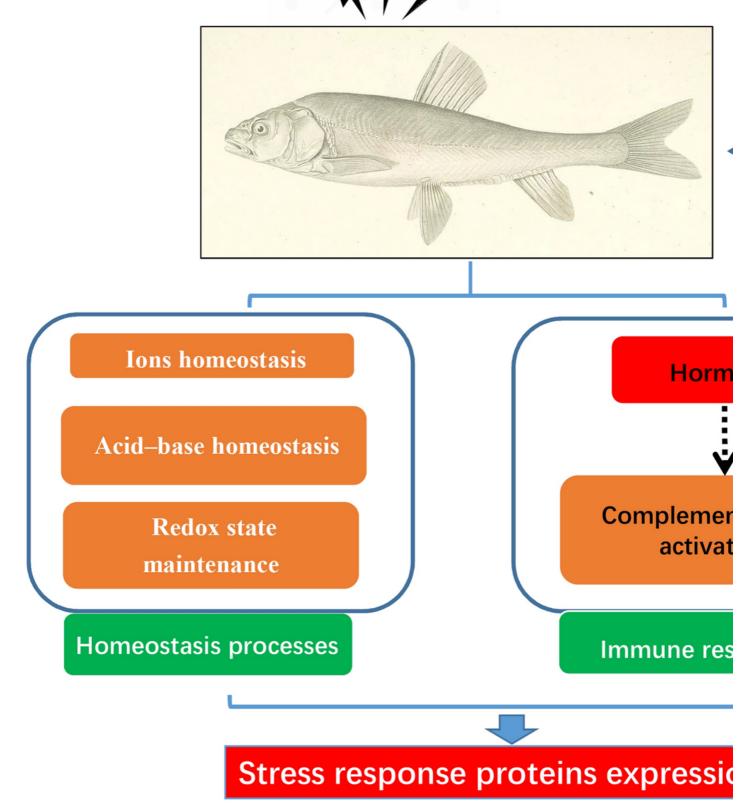
Bioresources and Bioprocessing 2022 9:104

Research Published on: 26 September 2022

Bioresources and Bioprocessing | Articles

Stress





## Biotransformation technology and high-value application of rapeseed meal: a review

Rapeseed meal (RSM) is an agro-industrial residue of increased functional biological value that contains high-quality proteins for animal feed. Due to the presence of antinutritional factors and immature devel...

Zhengfeng Yang, Zunxi Huang and Lijuan Cao

Bioresources and Bioprocessing 2022 9:103

Review Published on: 24 September 2022

> Full Text > PDF

## <u>Antifungal activity of silver/silicon dioxide nanocomposite on the response of faba bean plants</u> (*Vicia faba* L.) infected by *Botrytis cinerea*

Silicon (Si) and its nanomaterials could help plants cope with different negative effects of abiotic and/or biotic stresses. In this study, the antifungal role of silver/silicon dioxide nanocomposite (Ag/SiO<sub>2</sub>NC) ...

Zakaria A. Baka and Mohamed M. El-Zahed

Bioresources and Bioprocessing 2022 9:102

Research Published on: 19 September 2022

Bioresources and Bioprocessing | Articles





## An accessory enzymatic system of cellulase for simultaneous saccharification and co-fermentation

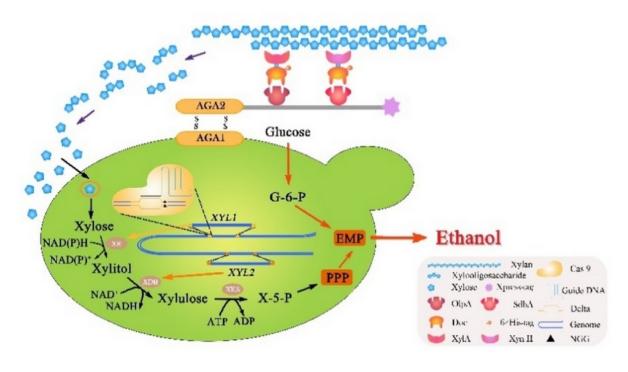
The enhanced hydrolysis of xylan-type hemicellulose is important to maximize ethanol production yield and substrate utilization rate in lignocellulose-based simultaneous saccharification and co-fermentation sy...

Han Liu, Xuxin Wang, Yanping Liu, Zhuoran Kang, Jiaqi Lu, Yutong Ye, Zhipeng Wang, Xinshu Zhuang and Shen Tian

Bioresources and Bioprocessing 2022 9:101

Research Published on: 19 September 2022

#### > Full Text > PDF



# <u>Effect of explant type (leaf, stem) and 2,4-D concentration on callus induction: influence of elicitor type (biotic, abiotic), elicitor concentration and elicitation time on biomass growth rate and costunolide biosynthesis in gazania (Gazania rigens) cell suspension cultures</u>

*Gazania rigens* (L.) Gaertn. (Asteraceae) is a medicinal plant with high ornamental potential and use in landscaping. The therapeutic potential of sesquiterpene lactones (SLs) as plant natural products for pharmac...

Huda E. Mahood, Virginia Sarropoulou and Thiresia-Teresa Tzatzani

Bioresources and Bioprocessing 2022 9:100

Research | Published on: 16 September 2022



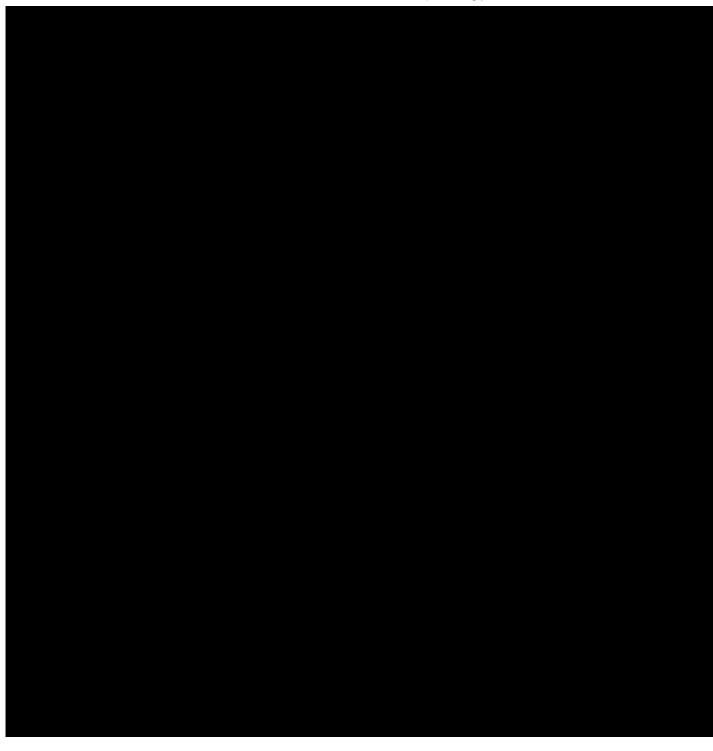
# In vitro assessment of multipotential therapeutic importance of *Hericium erinaceus* mushroom <u>extracts using different solvents</u>

Infectious disease is one of the major threats to humans and it is the second leading cause of death worldwide. Edible mushrooms have many nutritional and medicinal values to human health. The medicinal proper...

Waleed Bakry Suleiman, Reda M. Shehata and Ahmed M. Younis

Bioresources and Bioprocessing 2022 9:99

Research Published on: 16 September 2022



# Effective flow-through polishing strategies for knob-into-hole bispecific antibodies

Bispecific antibodies (bsAbs), though possessing great therapeutic potential, are extremely challenging to obtain at high purity within a limited number of scalable downstream processing steps. Complementary t...

Serene W. Chen, Kong Meng Hoi, Farouq Bin Mahfut, Yuansheng Yang and Wei Zhang

Bioresources and Bioprocessing 2022 9:98

Research Published on: 14 September 2022

# <u>Brazilian industrial yeasts show high fermentative performance in high solids content for corn</u> <u>ethanol process</u>

An imminent change in the world energy matrix makes it necessary to increase the production of renewable fuels. The United States and Brazil are the world's largest producers, but their production methods are ...

Thaís O. Secches, Carla F. Santos Viera, Thaynara K. E. Pereira, Victor T. O. Santos, Jade Ribeirodos Santos, Gonçalo A. G. Pereira and Marcelo F. Carazzolle

Bioresources and Bioprocessing 2022 9:97

Research Published on: 11 September 2022



# Induction of cellulase production by Sr<sup>2+</sup> in *Trichoderma reesei* via calcium signaling transduction

*Trichoderma reesei* RUT-C30 is a well-known high-yielding cellulase-producing fungal strain that converts lignocellulose into cellulosic sugar for resource regeneration. Calcium is a ubiquitous secondary messenger...

Ni Li, Yi Zeng, Yumeng Chen, Yaling Shen and Wei Wang

Bioresources and Bioprocessing 2022 9:96

Research Published on: 6 September 2022

> Full Text > PDF

# A novel pressed coal from citrus and cooking oil wastes using fungi

Nowadays renewable energy with low prices is a global target that has taken the attention to compare alternatives energy sources with fossil fuels. Therefore, this study was established to find suitable and su...

Mohamed S. Hasanin, Amr H. Hashem, Hassan M. Abu Hashish and Mohamed Abdelraof

Bioresources and Bioprocessing 2022 9:95

Research | Published on: 5 September 2022

> Full Text > PDF

# <u>Effect of chitooligosaccharides with a specific degree of polymerization on multiple targets in</u> <u>T2DM mice</u>

Chitooligosaccharides (COS) are found naturally in the ocean and present a variety of physiological activities, of which hypoglycemic action has attracted considerable research attention. This study aimed to a...

Jiangshan You, Mengyao Zhao, Shumin Chen, Lihua Jiang, Shuhong Gao, Hao Yin and Liming Zhao

Bioresources and Bioprocessing 2022 9:94

Research | Published on: 5 September 2022

> Full Text > PDF

# Light-driven progesterone production by InP-(M. neoaurum) biohybrid system

Progesterone is one of the classical hormone drugs used in medicine for maintaining pregnancy. However, its manufacturing process, coupled with organic reagents and poisonous catalysts, causes irreversible env...

Kun Liu, Feng-Qing Wang, Ke Liu, Yunqiu Zhao, Bei Gao, Xinyi Tao and Dongzhi Wei

Bioresources and Bioprocessing 2022 9:93

Research | Published on: 1 September 2022

# <u>Development of highly efficient whole-cell catalysts of *cis*-epoxysuccinic acid hydrolase by surface <u>display</u></u>

Bacterial *cis*-epoxysuccinic acid hydrolases (CESHs) are intracellular enzymes used in the industrial production of enantiomeric tartaric acids. The enzymes are mainly used as whole-cell catalysts because of the l...

Rui Zhou, Sheng Dong, Yingang Feng, Qiu Cui and Jinsong Xuan

Bioresources and Bioprocessing 2022 9:92

Research Published on: 29 August 2022

> Full Text > PDF

# <u>Efficient biodegradation of straw and persistent organic pollutants by a novel strategy using</u> <u>recombinant *Trichoderma reesei*</u>

Efficient biodegradation of lignocellulosic biomass needs a battery of enzymes targeting cellulose, hemicellulose, and lignin. In this study, recombinant *Trichoderma reesei* ZJ-09 with *Pycnoporus sanguineus* laccas...

Ying Xia and Xinda Lin

Bioresources and Bioprocessing 2022 9:91

Research Published on: 29 August 2022

> Full Text > PDF

# <u>Two strategies to improve the supply of PKS extender units for ansamitocin P-3 biosynthesis by</u> <u>CRISPR–Cas9</u>

Ansamitocin P-3 (AP-3) produced by *Actinosynnema pretiosum* is a potent antitumor agent. However, lack of efficient genome editing tools greatly hinders the AP-3 overproduction in *A. pretiosum*. To solve this probl...

Siyu Guo, Xueyuan Sun, Ruihua Li, Tianyao Zhang, Fengxian Hu, Feng Liu and Qiang Hua

Bioresources and Bioprocessing 2022 9:90

Research Published on: 29 August 2022

> Full Text > PDF

# <u>Characterization of pH-responsive high molecular-weight chitosan/poly (vinyl alcohol) hydrogel</u> <u>prepared by gamma irradiation for localizing drug release</u>

pH-sensitive hydrogels prepared by gamma irradiation find promising biological applications, partially, in the field of localized drug liberation. Herein, optimal conditions for fabricating high-molecular-weig...

#### Bioresources and Bioprocessing | Articles

Tu Minh Tran Vo, Thananchai Piroonpan, Charasphat Preuksarattanawut, Takaomi Kobayashi and Pranut Potiyaraj

Bioresources and Bioprocessing 2022 9:89

Research | Published on: 27 August 2022

> Full Text > PDF

# <u>Regiospecific C–H amination of (–)-limonene into (–)-perillamine by multi-enzymatic cascade</u> <u>reactions</u>

(-)-Limonene, one of cyclic monoterpenes, is an important renewable compound used widely as a key building block for the synthesis of new biologically active molecules and fine chemicals. (-)-Perillamine, as d...

Yue Ge, Zheng-Yu Huang, Jiang Pan, Chun-Xiu Li, Gao-Wei Zheng and Jian-He Xu

Bioresources and Bioprocessing 2022 9:88

Short report Published on: 26 August 2022

> <u>Full Text</u> > <u>PDF</u>

# <u>Vitamin combination promotes ex vivo expansion of NK-92 cells by reprogramming glucose</u> <u>metabolism</u>

Robust ex vivo expansion of NK-92 cells is essential for clinical immunotherapy. The vitamin B group is critical for the expansion and function of immune cells. This study optimized a vitamin combination by re...

Yan Fu, Yuying Chen, Zhepei Xie, Huimin Huang, Wen-Song Tan and Haibo Cai

Bioresources and Bioprocessing 2022 9:87

Research Published on: 26 August 2022

> Full Text > PDF

# <u>Bioeconomic production of high-quality chitobiose from chitin food wastes using an in-house</u> <u>chitinase from *Vibrio campbellii*</u>

Marine *Vibrio* species are natural degraders of chitin and usually secrete high levels of chitinolytic enzymes to digest recalcitrant chitin to chitooligosaccharides. This study used an endochitinase (*Vh*ChiA) from...

Reeba Thomas, Tamo Fukamizo and Wipa Suginta

Bioresources and Bioprocessing 2022 9:86

Research Published on: 20 August 2022

# <u>Ultrafine fully vulcanized natural rubber modified by graft-copolymerization with styrene and</u> <u>acrylonitrile monomers</u>

This research aims to modify ultrafine fully vulcanized powdered natural rubber (UFPNR) prepared by emulsion graftcopolymerization with styrene (St) and acrylonitrile (AN) monomers onto deproteinized natural ...

Krittaphorn Longsiri, Phattarin Mora, Watcharapong Peeksuntiye, Chanchira Jubsilp, Kasinee Hemvichian, Panagiotis Karagiannidis and Sarawut Rimdusit

Bioresources and Bioprocessing 2022 9:85

Research | Published on: 20 August 2022

> Full Text > PDF

## Characterization of a novel GH10 alkali-thermostable xylanase from a termite microbiome

The aim of the present study was to assess the biochemical and molecular structural characteristics of a novel alkali-thermostable GH10 xylanase (Xyl10B) identified in a termite gut microbiome by a shotgun met...

Maria Laura Mon, Rubén Marrero Díaz de Villegas, Eleonora Campos, Marcelo A. Soria and Paola M. Talia

Bioresources and Bioprocessing 2022 9:84

Research | Published on: 17 August 2022

> Full Text > PDF

## Production of cellulosic ethanol and value-added products from corn fiber

Corn fiber, a by-product from the corn processing industry, mainly composed of residual starch, cellulose, and hemicelluloses, is a promising raw material for producing cellulosic ethanol and value-added produ...

Yingjie Guo, Guodong Liu, Yanchun Ning, Xuezhi Li, Shiyang Hu, Jian Zhao and Yinbo Qu

Bioresources and Bioprocessing 2022 9:81

Review Published on: 13 August 2022

> Full Text > PDF

## Efficient synthesis 1,4-cyclohexanedicarboxaldehyde by an engineered alcohol oxidase

In this study, we selected and engineered a flavin adenine dinucleotide (FAD)-dependent alcohol oxidase (AOX) to produce 1,4cyclohexanedicarboxaldehyde (CHDA), an initial raw material for spiral compounds, fr...

Yaqi Cheng, Wei Song, Xiulai Chen, Cong Gao, Jia Liu, Liang Guo, Meng Zhu, Liming Liu and Jing Wu

Bioresources and Bioprocessing 2022 9:80

Research Published on: 13 August 2022

$\leftarrow Previous \qquad 1 \qquad 2 \qquad 3 \qquad Next \rightarrow$
How was your experience today?
Awful
Bad
СОК
Good
Great
Send feedback
Submit manuscript

#### Editorial Board

Sign up for article alerts and news from this journal

Affiliated with



*Bioresources and Bioprocessing* is associated with the <u>State Key Laboratory of Bioreactor Engineering</u>, East China University of Science and Technology.

# Annual Journal Metrics

#### 2022 Citation Impact

4.6 - 2-year Impact Factor6.1 - 5-year Impact Factor1.232 - SNIP (Source Normalized Impact per Paper)0.766 - SJR (SCImago Journal Rank)

#### 2022 Speed

6 days submission to first editorial decision for all manuscripts (Median) 80 days submission to accept (Median)

**2022 Usage** 897,118 downloads 131 Altmetric mentions

ISSN: 2197-4365 (electronic)

This journal is indexed by

- SCOPUS
- Science Citation Index Expanded
- Google Scholar
- CNKI
- DOAJ
- EBSCO Discovery Service
- EBSCO TOC Premier
- $\circ~$  OCLC WorldCat Discovery Service
- ProQuest Biological Science Database
- ProQuest Materials Science & Engineering Database
- ProQuest Natural Science Collection
- ProQuest SciTech Premium Collection
- ProQuest Technology Collection
- ProQuest-ExLibris Primo
- ProQuest-ExLibris Summon

Support and Contact	Terms and conditions		
Jobs	Privacy statement		
Language editing for authors	<u>Accessibility</u>		
Scientific editing for authors	<u>Cookies</u>		
Leave feedback			

Follow SpringerOpen



By using this website, you agree to our <u>Terms and Conditions</u>, <u>Your US state privacy rights</u>, <u>Privacy statement</u> and <u>Cookies</u> policy. <u>Your privacy choices/Manage cookies</u> we use in the preference centre.

#### **SPRINGER NATURE**

© 2023 BioMed Central Ltd unless otherwise stated. Part of Springer Nature.

# RESEARCH

**Open Access** 

# Utilization of durian seed for *Monascus* fermentation and its application as a functional ingredient in yogurt

Ignatius Srianta, Indah Kuswardani, Susana Ristiarini, Netty Kusumawati, Laura Godelive and Ira Nugerahani<sup>\*</sup>

#### Abstract

As a widely consumed fermented milk product, yogurt undergoes constant development to increase its functional properties. Monascus purpureus-fermented durian seed, which has been proven to possess antioxidative properties, has the potential to improve yogurt properties. This study aimed to analyze the use of Monascus-fermented durian seed (MFDS) as a functional ingredient in yogurt and its effect on physicochemical properties, lactic acid bacteria (LAB) count, antioxidative properties, and consumer acceptability of set-type yogurt during refrigeration. Changes in physicochemical properties, including color, pH, titratable acidity, syneresis, LAB count, total phenolic content (TPC), and antioxidant activity were evaluated at 7-day intervals during 14 days of refrigerated storage (4 °C). Sensory evaluations were carried out for freshly made samples after 7 days of storage. The results showed that the addition of MFDS to yogurt gave significant effects on some of the parameters measured. Yogurt with added MFDS powder produced a more red color ( $L = 88.55 \pm 1.28$ ,  $a^* = 2.63 \pm 0.17$ ,  $b^* = 11.45 \pm 1.15$ ,  $c = 11.75 \pm 1.15$ ,  $H = 77.00 \pm 0.64$ ), reached the highest TPC (2.21  $\pm$  0.46 mg/GAE g), antioxidant activity (0.0125  $\pm$  0.0032 mg GAE/g), and syneresis (5.24  $\pm$  0.51%) throughout 14 days of storage. The addition of MFDS only gave a slight difference to pH and titratable acidity, while no significant difference was made for LAB count. For sensory evaluation, the addition of MFDS, particularly the ethanol extract, to yogurt was well-liked by panelists. Citrinin content in MFDS yogurt can be decreased under the limits set. Overall, the addition of MFDS has a high potential of improving yogurt properties, particularly its antioxidative properties.

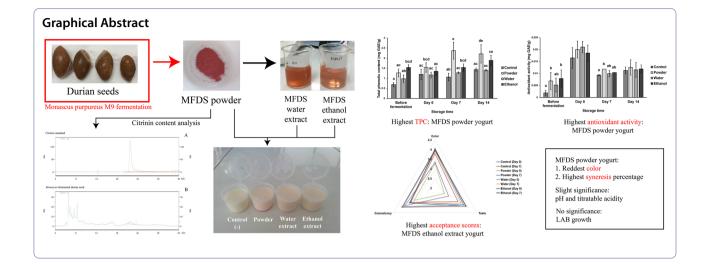
**Keywords:** *Monascus*-fermented durian seed, Yogurt, Antioxidant, Phenolic content, *Monascus purpureus*, Refrigerated storage

\*Correspondence: ira@ukwms.ac.id

Department of Food Technology, Faculty of Agricultural Technology, Widya Mandala Catholic University Surabaya, Jalan Dinoyo 42-44, Surabaya 60295, Indonesia



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.



#### Introduction

Yogurt is a fermented dairy product usually produced with Streptococcus thermophilus and Lactobacillus delbrueckii subsp. bulgaricus. Yogurt is rich in activity probiotics and nutritional compounds, such as calcium, protein, and potassium (Gahruie et al. 2015; Liu and Lv 2018). Lactic acid bacteria (LAB) in yogurt are credited with improving digestion, harmful bacteria growth suppression in the gastrointestinal tract, the bioavailability of milk constituents, alleviation of lactose intolerance, cancer suppression, and hypocholesterolemic effects (Shiby and Mishra 2013; Aryana and Olson 2017). Owing to its functional properties, yogurt has been widely consumed for thousands of years and undergoes constant development to improve its health benefits. One of the most common ways to improve yogurt quality is by adding functional ingredients, such as essential oils (Comunian et al. 2017), fruits, flowers (Chouchouli et al. 2013; Marchiani et al. 2015; Sah et al. 2016; Liu and Lv 2018; Dimitrellou et al. 2020), green tea and coffee powders (Dönmez et al. 2017), and Monascus-fermented products (Jeon et al. 2011; Abdel-Raheam et al. 2019).

Monascus-fermented products are substrates fermented by Monascus sp., typically from the three species: Monascus purpureus, Monascus pilosus, and Monascus ruber. The most notable Monascus product is red mold rice or commonly known in Indonesia as angkak, which is a Monascus-fermented rice product. These products have been commonly used as food colorants and dietary material due to the secondary metabolites produced during fermentation: Monascus pigments, monacolin K,  $\gamma$ -aminobutyric acid (GABA), and dimerumic acid (Lee and Pan 2012). These compounds have been reported to have bioactivities, such as anti-inflammatory, anticancer, antimicrobial, antiobesity, antidiabetic, and antihypercholesterolemia (Srianta et al. 2021). Jeon et al. (2011) reported on the addition of Monascus sp. and Lactobacillus sp. fermented Chinese yam (Dioscorea batatas) powder in yogurt and found that the addition increased total phenolic content, 2,2-diphenyl-1-picrylhydrazyl (DPPH) scavenging activity, reducing power, angiotensin-converting enzyme (ACE) inhibitory activity, and GABA content of yogurt in comparison to the control. Abdel-Raheam et al. (2019) also reported that the addition of red, orange and yellow pigments from Monascus ruber fermentation in yogurt received high acceptability scores on texture, odor, color, taste, and overall acceptability. These results show the potential of Monascusfermented products to be added as a yogurt ingredient, especially with substrates other than rice.

Durian is one of the many tropical fruits largely produced in Indonesia. Waste from durian production can go as high as 70% which consists of shells and seeds (Purnomo et al. 2016). Srianta et al. (2012), Subianto et al. (2013), and Nugerahani et al. (2017) reported that Monascus purpureus fermentation on durian seeds is possible and has been proven to exhibit antioxidant, antihypercholesterol, and antidiabetic activities due to Monascus pigments, phenolics, and monacolin K. These compounds have different polarities, which could affect MFDS bioactivity according to the extraction solvent (Subianto et al. 2013). This raises the potential of Monascus-fermented durian seeds (MFDS) of various forms to be sustainable and functional ingredients for food products, especially yogurts. This study aimed to analyze the use of MFDS as a functional ingredient in yogurt and its effect on physicochemical properties, LAB count, antioxidative

properties, and consumer acceptability of set-type yogurt during refrigeration.

#### **Materials and methods**

#### Monascus-fermented durian seed preparation

The durian seeds of the Petruk variety were provided by a local durian product seller in Surabaya, Indonesia. The seeds were washed clean, sorted, and kept in the freezer (Sharp, Japan) at -20 °C before use. The seeds were thawed for 30 min (30 °C), washed, weighed, and heated with 5% (w/v) Ca(OH)<sub>2</sub> solution (1:1) for 10 min at 85-90 °C. The seeds were washed and separated from their skin, diced into smaller pieces (1 cm<sup>3</sup>), weighed (50 g/batch) inside 250 mL titration flasks, sterilized with an autoclave (121 °C, 15 lbs/inch<sup>2</sup>, 15 min, Hirayama, Saitama, Japan), and cooled to 30 °C. The sterilized durian seeds were inoculated with 5% (v/w) Monascus purpureus M9 (NCBI Accession Number: HM188425.1) and left to ferment for 14 days at 30 °C to obtain MFDS. The MFDS was then dried in an oven for 24 h at 45 °C (Binder, Germany), ground, and sieved (80 mesh) to obtain MFDS powder. MFDS powder would then be analyzed for citrinin content.

MFDS powder was then extracted with two different solvents, water and ethanol. MFDS that had been weighed were extracted with sterile distilled water (1:50 w/v) using a shaking water bath (LabTech, Hopkinton, Massachusetts) at 100 rpm, 40 °C, for 1 h and strained using a vacuum pump to obtain the MFDS water extract. The MFDS ethanol extract was made by extracting MFDS powder with ethanol (1:50 w/v) using the Soxhlet method for 2.5 h. The extract was then evaporated with a rotary evaporator (70 °C, 50 rpm, Buchi, Switzerland) and rehydrated with sterile distilled water. Both MFDS extracts were pasteurized for 30 min at 70 °C in a water bath (Faithful, Hebei, China) and stored at 4 °C.

#### Yogurt preparation

Commercial ultra-high temperature sterilized full cream milk (Ultrajaya, Indonesia) was purchased from a local market. Granulated sugar (10% w/v) and skimmed milk powder (2.2% w/v) were added to the milk and mixed thoroughly. The mixture was pasteurized at 90 °C for 5 min above a Bunsen burner fire. Gelatin (0.8% w/v) and MFDS treatments as illustrated in Table 1 were added to the mixture at 80 °C. After it was homogenously mixed, the mixture was taken off the fire and cooled in an ice bath to 41 °C. Commercial yogurt starter (a mixture of

Page 3 of 14

Ingredient	Treatment				
	Control	Powder	Water	Ethanol	
Full cream milk (mL)	700				
Granulated sugar (w/v) <sup>a</sup>	10%				
Skimmed milk powder (w/v)	2.2%				
Gelatin (w/v) <sup>a</sup>	0.8%				
MFDS powder (w/v) <sup>b</sup>	0	0.15%	0	0	
MFDS water extract (v/v) <sup>b</sup>	0	0	7.5%	0	
MFDS ethanol extract (v/v) <sup>b</sup>	0	0	0	7.5%	
Water (v/v) <sup>b</sup>	7.5%	6.35%	0	0	
Starter culture (w/v) <sup>c</sup>	0.3%				

<sup>a</sup> Calculated based on the milk volume

 $^{\rm b}$  Calculated based on the milk, sugar, skimmed milk powder, and gelatin total volume

<sup>c</sup> Calculated based on the total volume of the mixture after MFDS treatment

*Streptococcus thermophilus, Lactobacillus bulgaricus*, and *Lactobacillus acidophilus*, Lallemand, France) of 0.3% (w/v) was inoculated and mixed into the yogurt mixture. The mixture was poured into sterile plastic cups with some put aside for pH analysis. Fermentation was conducted at 42 °C for 4 h to obtain MFDS yogurt. Color, syneresis, titratable acidity, pH, total LAB count, antioxidant activity, phenolic content, and sensory evaluation were determined after fermentation. The yogurt samples were cooled at 4 °C for 14 days and underwent analysis of the parameters mentioned at 7-day intervals.

#### Physicochemical analysis

Color parameters of lightness ( $L^*$ ), redness ( $a^*$ ), and yellowness ( $b^*$ ) were determined by using a color reader (Konica Minolta, Japan). Chroma and hue parameters were further derived from  $a^*$  and  $b^*$  values based on Eqs. (1) and (2):

$$Chroma = \sqrt{a^{*2} + b^{*2}} \tag{1}$$

$$Hue = \arctan(b^*/a^*) \tag{2}$$

Syneresis analysis was conducted according to Amatayakul et al. (2006). The yogurt samples inside the plastic cups were weighed to obtain the initial weights before the cups were tilted to 45° for whey extraction with a pipette and filtration paper. After extraction, the yogurt samples were weighed to obtain the final weight. Syneresis percentage was calculated according to Eq. (3):

(3)

%Syneresis = 
$$\{(initial weight - final weight)/(initial weight - empty cup weight)\} \times 100\%$$

The pH of the samples was determined by using a pH meter (Xylem Analytics, Germany). The titratable acidity (lactic acid) was determined according to the methodology suggested by Widagdha and Nisa (2015). Standardized NaOH was used to titrate 10 mL of the sample (previously diluted tenfold) with added phenolphthalein indicator until the solution reaches a stable pink color. The results were expressed as total lactic acid percentages.

#### **Microbiological analysis**

The counts of LAB were done in duplicate using de Man, Rogosa and Sharpe agar and incubated for 48 h at 37 °C. The results were expressed as colony-forming units per milliliter of yogurt (CFU mL<sup>-1</sup>).

#### Antioxidant activity

Determination of the samples' antioxidant activity was done with the DPPH assay according to Subianto et al. (2013). In preparation for the assay, distilled water was added to the samples (1:1), centrifuged for 30 min at 5000 rpm, and filtered with Whatman number 42 filter paper. The supernatants were diluted fivefold to obtain the samples for DPPH assay. The samples and the DPPH solution reacted for 30 min and the absorbance (A) was measured at 517 nm in a UV–Vis spectrophotometer (Shimadzu, Japan). The Gallic acid standard curve was conducted with methanol as the solvent. The samples' antioxidant activity was expressed in milligrams of Gallic acid equivalent per gram of yogurt (mg GAE·g<sup>-1</sup>) and calculated according to Eq. (4):

%Inhibition =  $\{(A \text{ control} - A \text{ sample})/(A \text{ control})\} \times 100\%.$ (4)

#### Phenolic content

The preparation of the samples was carried out using the same procedure as the samples for the DPPH assay. The samples (0.1 mL) were reacted to 0.5 mL Folin Ciocalteu reagent inside a 10 mL volumetric flask, homogenized, then 1.5 mL of 20% Na<sub>2</sub>CO<sub>3</sub> and distilled water were added. After homogenization, the solution was put aside for 30 min at room temperature and the absorbance was measured at 750 nm. The Gallic acid standard curve was conducted with distilled water as the solvent. The results were expressed in milligrams of Gallic acid equivalent per gram of yogurt (mg GAE·g<sup>-1</sup>).

#### Sensory evaluation

Fifty untrained students of Widya Mandala Surabaya Catholic University who have basic sensory knowledge

of yogurt products were instructed to evaluate the color, taste, and consistency of yogurt samples. A hedonic 5-point scale was used, in which '5' represents 'most like', '4' represents 'like', '3' represents 'neutral', '2' represents 'dislike', and '1' represents 'most dislike' (Setyaningsih et al. 2010).

#### **Citrinin content**

Citrinin content analysis of MFDS was carried out according to Li et al. (2003). MFDS (0.5 g) was extracted with 20 mL of toluene—ethyl acetate—formic acid reagent (7:3:1, v/v/v) and disintegrated for 10 min in an ultrasonic disintegrator. The extract and its container were weighed pre and post-sonication and added with the complex extract reagent to make up for the deficiency. The residue was extracted twice with 15 mL of the reagent and was combined with the initial extract to be centrifuged at 3000 rpm for 20 min. The upper layer of the extract was evaporated and the residue was dissolved in 30 mL of methanol for citrinin determination by high-performance liquid chromatography (HPLC) after filtration.

The HPLC system (Hewlett-Packard 1100, USA) was used for citrinin determination. A reversed phase column (Eclipse XDB C<sub>18</sub>, 4.6 mm × 250 mm, 5 µm) was used after being thermostated at 28 °C in a column oven. The system included a fluorescence detector with an excitation wavelength of 331 nm/500 nm. The samples were eluted with acetonitrile-acidified water (pH 2.5, 35:65, v/v) at a flow rate of 1 mL/min. The detection limit was 50 µg/L.

Citrinin confirmation was done with a liquid chromatography-mass spectrometry (LC-MS) system (Waters ZMD 4000), with an inlet temperature of 120 °C and a dissolvent temperature of 25 °C. The capillary voltage was 3.78 kV and the molecular weights were 100 to 500 Da. The same mobile phase, column, and employed flow rate were used.

#### Statistical analysis

The data were analyzed using IBM SPSS Statistics (version 19.0). The added MFDS and storage time of the yogurt samples were the two factors that could affect the parameters tested. Two-way ANOVA was conducted and continued with post hoc pairwise comparisons using Tukey's HSD test (p < 0.05) when the effects were significant.

# **Results and discussion**

#### **Color analysis**

For all food products, including yogurt, color is one of the visual attributes which has a part in influencing consumers' preferences. Data of each yogurt sample's  $L^*$ 

Color parameter	Storage time (day)	Treatment			
		Control	Powder	Water	Ethanol
L	Before fermentation	$89.32 \pm 0.93^{abcd}$	88.12±0.53ª	$88.10 \pm 2.14^{a}$	$89.60 \pm 1.11^{abcd}$
	0	89.32 ± 1.63 <sup>abcd</sup>	$89.45\pm0.93^{abcd}$	$90.63 \pm 0.38^{cde}$	$89.93 \pm 1.94^{\text{bcde}}$
	7	$90.68 \pm 0.77^{de}$	$88.65\pm0.89^{ab}$	89.03 ± 1.53 <sup>abc</sup>	$91.35 \pm 0.94^{e}$
	14	$90.00 \pm 1.18^{bcde}$	$88.55 \pm 1.28^{ab}$	$90.40\pm0.57^{cde}$	$90.88 \pm 0.53^{de}$
a*	Before fermentation	$-0.33 \pm 0.54^{a}$	$2.38 \pm 0.10^{fg}$	$1.25 \pm 0.16^{e}$	$-0.03 \pm 0.22^{ab}$
	0	$0.17 \pm 0.23^{abc}$	$2.08\pm0.34^{\rm f}$	$1.30 \pm 0.70^{e}$	$0.18 \pm 0.12^{abc}$
	7	$0.73 \pm 0.12^{d}$	$2.25\pm0.05^{\rm fg}$	$1.52 \pm 0.17^{e}$	$0.53\pm0.26^{cd}$
	14	$0.12 \pm 0.24^{abc}$	$2.63 \pm 0.17^{9}$	$1.33 \pm 0.26^{e}$	$0.38\pm0.50^{bcd}$
<i>b</i> *	Before fermentation	$11.13 \pm 0.62^{bA}$	$9.68 \pm 0.39^{aA}$	$10.22 \pm 0.59^{abA}$	$10.70 \pm 0.98^{ab}$
	0	11.98±0.39 <sup>bB</sup>	$10.90 \pm 0.74^{aB}$	10.90±1.27 <sup>aAB</sup>	$11.18 \pm 0.93^{a}$
	7	$12.08 \pm 0.39^{B}$	$11.62 \pm 0.82^{B}$	$11.82 \pm 0.63^{B}$	$11.45 \pm 0.89$
	14	$11.58 \pm 0.15^{AB}$	$11.45 \pm 1.15^{B}$	$11.53 \pm 0.90^{AB}$	$11.43 \pm 0.72$
С	Before fermentation	$11.15 \pm 0.60^{A}$	$9.98 \pm 0.39^{A}$	$10.32 \pm 0.59$	$10.70 \pm 0.98$
	0	11.98±0.39 <sup>BC</sup>	$11.87 \pm 1.82^{B}$	$11.30 \pm 1.84$	$11.25 \pm 1.02$
	7	$12.08 \pm 0.39^{\circ}$	$11.82 \pm 0.82^{B}$	$11.92 \pm 0.63$	$11.45 \pm 0.89$
	14	$11.58 \pm 0.15^{AB}$	$11.75 \pm 1.15^{AB}$	11.63±0.90	$11.43 \pm 0.72$
Н	Before fermentation	$87.48 \pm 2.27^{de}$	$76.18 \pm 0.18^{a}$	$83.03 \pm 0.90^{\circ}$	$90.27 \pm 1.23^{f}$
	0	$88.75\pm0.79^{\text{def}}$	$78.70 \pm 1.86^{b}$	$83.37 \pm 3.14^{\circ}$	$88.88 \pm 0.76^{ef}$
	7	$86.52 \pm 0.68^{d}$	$78.98 \pm 0.87^{b}$	$82.67 \pm 1.15^{\circ}$	$87.42 \pm 1.09^{de}$
	14	$89.43 \pm 1.18^{ef}$	$77.00 \pm 0.64^{ab}$	$83.32 \pm 1.74^{c}$	$88.20\pm2.36^{def}$

Table 2 Changes in lightness (L), redness (a\*), yellowness (b\*), chroma (c), and hue (H) of yogurt samples during storage (4 °C)

 $^{\rm a-\ f}$  Mean values with different letters are significantly different (p < 0.05)

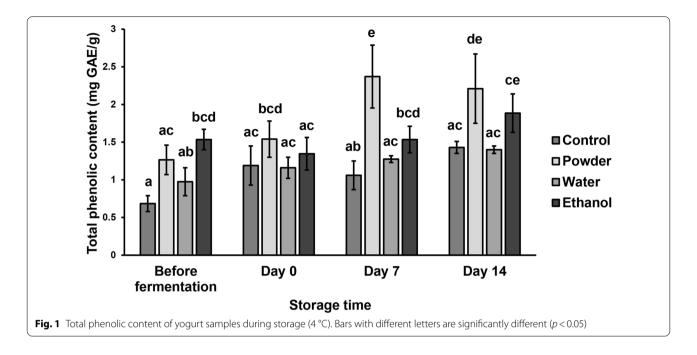
<sup>A, B</sup>Mean values in the same column with different letters are significantly different (p < 0.05)

(lightness), a\* (redness), b\* (yellowness), c (chroma), and H (hue) are shown in Table 2. Treatments of MFDS forms had a significant difference in the  $L^*$ ,  $a^*$ ,  $b^*$ , and h values of yogurt, particularly in the  $a^*$  value. MFDS has a bright red color from the pigments produced by Monascus purpureus during fermentation e.g. the yellow pigments: monascin and ankaflavin, orange pigments: rubropunctatin and monascorubrin, and the red pigments: rubropunctamine and monascorubramine (Feng et al. 2012). These added pigments would give a significant effect on the yogurt color in comparison to the control. Yogurt with added MFDS powder also produced the highest  $a^*$  values and the lowest H values throughout storage, while yogurt with added MFDS water extract produced the second highest  $a^*$  values. Overall, yogurt with added MFDS powder produced a redder color compared to other yogurt samples.

The extraction process of MFDS could decrease the color intensity of the MFDS water and ethanol extracts by losing some of the pigments during extraction. Some pigments may have been degraded from the high temperature of extraction and pasteurization. Most *Monascus* pigments are unstable outside of 30–60 °C (Feng et al. 2012). Also, the pigments may have not been fully

extracted from the MFDS powder due to the solvent used. Srianta et al. (2019) found that MFDS extracted using ethanol:water solvent with a ratio of 7:3 produced higher red pigment content compared to extraction using a 10:0 ratio solvent. Meanwhile, MFDS powder was added straight into the yogurt mixture, thus most of the pigments were able to be incorporated into the yogurt without any further degradation from the extraction process.

Yogurt with added ethanol extract was less red in comparison to the yogurt with added water extract. The six main *Monascus* pigments are ethanol soluble, but reactions with -COOH or NH<sub>3</sub> groups of amino acids would produce water-soluble derivative pigments (Feng et al. 2012). Srianta et al. (2012) found more water-soluble yellow, orange, and red pigments in MFDS, which might be due to the amino acid content in durian seeds as a source of NH<sub>3</sub>. A study by Mirhosseini et al. (2013) found that leucine, lysine, aspartic acid, glycine, alanine, glutamic acid, valine, proline, serine, threonine, isoleucine, and phenylalanine were the most abundant amino acids in the chemical structure of durian seed gum. More pigments were extracted with distilled water, resulting in yogurt with a redder color than using the ethanol extract.



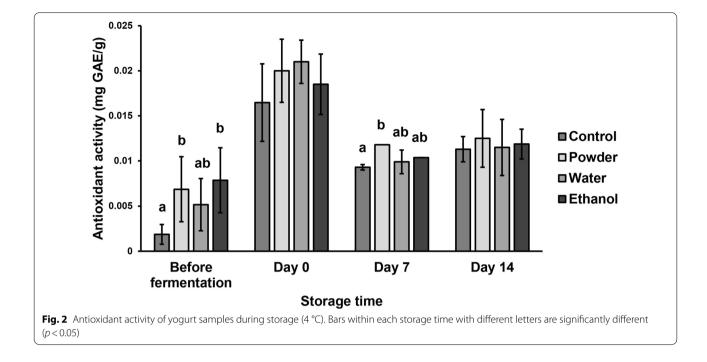
#### **Total phenolic content**

The total phenolic content data of all yogurt samples is shown in Fig. 1. Yogurt with added MFDS powder reached the highest total phenolic content (TPC) throughout 14 days of storage, with  $1.54\pm0.24$  mg GAE/g on day 0,  $2.37\pm0.42$  mg GAE/g on day 7, and  $2.21\pm0.46$  mg/GAE g on day 14. Yogurt with added MFDS ethanol extract reached the second highest TPC ( $1.35\pm0.21$ ,  $1.53\pm0.17$ , and  $1.89\pm0.25$  mg GAE/g on day 0, day 7, and day 14, respectively), while the addition of MFDS water extract did not affect TPC in comparison to the control. MFDS water extract and ethanol extracts were reported to have TPC as high as 3.58 mg GAE/g and 3.61 mg GAE/g, respectively (Srianta et al. 2014).

Durian seeds contain natural phenolics and exhibit antioxidant activity (Subianto et al. 2013). According to Ramli et al. (2021), freeze-dried and rotary evaporated durian seeds had TPC of 4.46 mg GAE/mL and 29.52 mg GAE/mL, respectively. The TPC of durian seeds from previous studies being higher than MFDS extracts might be due to differences in the extraction process and the variety of durian used. Another study by Srianta et al. (2013) compared durian seed water and ethanol extract with MFDS water and ethanol extract. MFDS water extract (589.8 µg GAE/mL) and ethanol extract (473.3 µg GAE/mL) exhibited higher TPC than durian seed water extract (325.8 µg GAE/mL) and ethanol extract (132.0 µg GAE/mL). Monascus sp. fermentation can further increase the TPC of substrates and their bioavailability by releasing enzymes including amylases, cellulases, esterases, tannases, and glucosidases to break down cell walls and facilitate phenol extraction (Suraiya et al. 2018; Zhihao et al. 2022). These enzymes can also break bonds between phenolic compounds and other groups such as polysaccharides, lipids, and amines (Cheng et al. 2022; Wang et al. 2018). *Monascus* sp. can also produce micromolecular phenolic compounds by itself (Bei et al. 2017).

The addition of MFDS powder in yogurt resulted in the highest TPC of all the samples, which is due to fewer processing steps of MFDS before being incorporated into the yogurt. A larger amount of phenolic compounds in the MFDS powder could have been preserved and thus able to be detected during the TPC analysis. Ethanol and water extracts of MFDS underwent a few more heat processing steps and would have lost more phenolics due to degradation. Based on the TPC of the yogurt samples, it is observed that phenolics in MFDS were more ethanol soluble rather than water soluble. Abd Razak et al. (2015) also found similar results where methanol extracts of rice bran fermented with *Rhizopus oligosporus* and *Monascus purpureus* had higher amounts of TPC compared to the water extracts.

TPC of yogurt with added MFDS powder and ethanol extract experienced an increase throughout 14 days of storage. The addition of MFDS powder increased the TPC of yogurt from  $1.54\pm0.24$  to  $2.21\pm0.46$  mg/GAE g during 14 days of storage, while MFDS ethanol extract increased the TPC from  $1.35\pm0.21$  to  $1.89\pm0.25$  mg GAE/g. Aside from the phenolic compounds present in MFDS, the degradation of milk proteins during fermentation by LAB could release phenolic amino acids and nonphenolic compounds which could have interfered



during TPC analysis (Shori 2013; Baba et al. 2014). Similar results were found by Szołtysik et al. (2020), in which the phenolic compounds (anthocyanins, favan-3-ols, ellagitannins, and flavonols) in yogurt with added *Rosa spinosissima* extract experienced a slight elevation after 14 days of storage. Yogurt with added *Azadirachta indica* also experienced an increase in TPC from  $38.5 \pm 7.2 \ \mu g$  GAE/mL on day 14 to  $74.9 \pm 6.2 \ \mu g$  GAE/mL by day 28 (Shori and Baba 2013).

#### Antioxidant activity

The antioxidant activity of yogurt samples was determined based on their DPPH scavenging capacities and are shown in Fig. 2. On day 0, yogurt with added MFDS water extract reached the highest antioxidant activity  $(0.0210 \pm 0.0035 \text{ mg GAE/g})$  while the control had the lowest activity  $(0.0165 \pm 0.0043 \text{ mg GAE/g})$ . Through day 7 and day 14, yogurt with added MFDS powder produced the highest antioxidant activity (0.0118 and  $0.0125 \pm 0.0032$  mg GAE/g respectively), while the control reached the lowest values with  $0.0093 \pm 0.0003$  mg GAE/g on day 7 and  $0.113 \pm 0.0014$  mg GAE/g on day 14. The addition of any form of MFDS increased the antioxidant activity of yogurt, which was due to the bioactive compounds produced by M. purpureus which are Monascus pigments, GABA, and monacolins. Srianta et al. (2017) reported that Monascus purpureus M9 pigments monapilol B and rubropunctamine were the main antioxidants of Monascus-fermented products among the 12 pigments detected (rubropunctatin, monascorubrin, rubropunctamine, monascorubramine, monascin, ankaflavin, xanthomonascin A, xanthomonascin B, yellow II, and monapilol B). The two pigments had a high correlation with the DPPH radical scavenging activity. Tan et al. (2018) found that water-soluble yellow *Monascus* pigments exhibited DPPH and 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS<sup>+</sup>) radical scavenging activity, reaching up to 86.33% inhibition from 1 mg/ mL sample and 99.95% from 0.25 mg/mL respectively. Other secondary metabolites such as GABA, monacolin K, and dihydromonacolin MV also contribute to DPPH radical scavenging (Srianta et al. 2014). Phenolics from *Monascus*-fermented oats were also reported to have a high correlation with DPPH and ABTS<sup>+</sup> radical scavenging activity (Bei et al. 2017).

After fermentation, the antioxidant activities of all the yogurt samples were significantly higher than the initial yogurt mixture. The increase in antioxidant activity of the yogurt mixture after fermentation was due to LAB activity. Using the thiobarbituric acid method, Kim et al. (2005) found that *Lactobacillus bulgaricus* and *Lactobacillus acidophilus* exhibit antioxidant activity (81.30% and 65.32% respectively). They can demonstrate radical scavenging activity, inhibitory activity towards lipid peroxidation, show strong reducing power, and produce proteolytic enzymes which help release antioxidative milk peptides (Virtanen et al. 2007; Zhang et al. 2011; Aloglu and Oner 2011). Gjorgievski et al. (2014) found that yogurt fermented with *Streptococcus thermophilus* and *L. bulgaricus* mix culture and *L. acidophilus* monoculture

also exhibited antioxidant activity by DPPH radical scavenging (52.44% and 63.99% respectively).

The antioxidant activity of the yogurt samples then significantly decreased by day 7 and stayed stagnant between day 7 and day 14. This data has a negative correlation with the TPC of yogurt samples throughout storage time, in which yogurt with added MFDS powder and ethanol extract had increasing TPC up until day 14. This suggests that the phenolics detected in yogurt with added MFDS did not have DPPH radical scavenging properties. Bei et al. (2017) found that the bound phenolic fraction of fermented oats with Monascus anka, mainly ferulic acid, showed greater ABTS<sup>+</sup> scavenging activity than DPPH radical scavenging activity. Abd Razak et al. (2015) reported that TPC and radicalscavenging activity of rice bran fermented with R. oligosporus and M. purpureus were poorly correlated for the water and methanol extracts, which might be due to many factors such as the concentration and chemical structures of the phenolics detected. This result suggests that other antioxidant compounds in MFDS might have stronger DPPH scavenging activity than the phenolics detected i.e. pigments, monacolins, and GABA (Srianta et al. 2014; Suraiya et al. 2018). These compounds would have been degraded during storage, thereby decreasing the antioxidant activity of yogurt samples. Similar results have been observed by Chouchouli et al. (2013), in which the antiradical capacities of grape seed-fortified full-fat yogurt decreased during 32 days of storage time  $(\text{from } 1487.4 \pm 38.2 - 1567 \pm 52.8 \text{ mg TE}/100 \text{ g on day 1 to})$  $234.8 \pm 20.6 - 440.5 \pm 40.3$  mg TE/100 g by day 32).

#### pH and titratable acidity

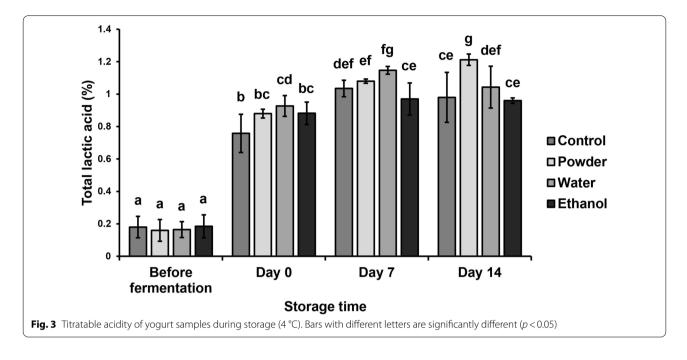
Changes in yogurt pH added with different MFDS forms are shown in Table 3. pH of the unfermented yogurt mixture decreased significantly after fermentation due to the formation of acids by LAB. pH data from yogurt with added MFDS powder experienced the biggest reduction during storage time from  $4.492 \pm 0.093$  on day 0 to  $4.175 \pm 0.087$  by day 14. Yogurt with added water extract reached the lowest pH on day 0 and day 7, while MFDS powder yogurt has the lowest pH by day 14. These results were in line with the titratable acidity data shown in Fig. 3. The majority of acids found in yogurt are lactic acids, produced by LAB by converting lactose in milk (Chen et al. 2017). The total lactic acid percentage of all yogurt mixture samples increased significantly after fermentation and increased gradually throughout storage time, which is in line with the pH data. The highest total acid percentage was reached by MFDS powder yogurt by day 14. Overall, the addition of MFDS forms to yogurt only gave a slight effect on the pH and titratable acidity in comparison to the control. Darwish et al. (2017) and Melani et al. (2021) found that the addition of angkak (Monascus-fermented rice/red mold rice) did not give a significant effect on the pH value and acidity of yogurt and goat milk kefir respectively. Pyo and Song (2009) found that the addition of Monascus-fermented soybean decreased the pH and increased the acidity of yogurt, while Jeon et al. (2011) found that the pH of yogurt increased after the addition of Monascus-fermented Chinese yam. Different substrates of Monascus-fermentation will produce different amounts and types of compounds which could affect the characteristics of each Monascusfermented product.

Parameter	Treatment	Storage time (day)				
		Before fermentation	0	7	14	
рН	Control	$6.397 \pm 0.048^{\circ}$	$4.468 \pm 0.068^{\text{bAB}}$	$4.291 \pm 0.096^{aAB}$	$4.289 \pm 0.130^{aAB}$	
	Powder	$6.415 \pm 0.034^{\circ}$	$4.492 \pm 0.093^{bB}$	$4.275 \pm 0.111^{aAB}$	$4.175 \pm 0.087^{aA}$	
	Water	$6.396 \pm 0.032^{\circ}$	$4.353 \pm 0.028^{bA}$	$4.215 \pm 0.045^{aA}$	$4.249 \pm 0.097^{aAB}$	
	Ethanol	$6.389 \pm 0.033^{b}$	$4.473 \pm 0.105^{aAB}$	$4.390 \pm 0.089^{aB}$	$4.373 \pm 0.087^{aB}$	
Syneresis (%)	Control	-	$2.22 \pm 0.63^{A}$	$2.42 \pm 0.60^{A}$	$2.56\pm0.68^{\text{A}}$	
	Powder	-	$3.29 \pm 0.43^{aB}$	$3.10 \pm 0.27^{aA}$	$5.24 \pm 0.51^{bB}$	
	Water	-	$2.83\pm0.82^{AB}$	$2.67\pm0.50^{A}$	$3.60\pm0.60^{AB}$	
	Ethanol	-	$2.59 \pm 0.46^{A}$	$2.68 \pm 0.60^{A}$	$2.92\pm0.69^{\text{A}}$	
Total plate count (CFU/mL)	Control	$2.65 \times 10^{7a}$	6.27 × 10 <sup>9b</sup>	3.36 × 10 <sup>10bc</sup>	$1.96 \times 10^{11c}$	
	Powder	3.05 × 10 <sup>6a</sup>	6.22 × 10 <sup>9a</sup>	$2.04 \times 10^{10a}$	$1.90 \times 10^{7a}$	
	Water	1.34 × 10 <sup>6a</sup>	7.82 × 10 <sup>10b</sup>	$4.84 \times 10^{10b}$	$1.16 \times 10^{9ab}$	
	Ethanol	4.00 × 10 <sup>6a</sup>	$1.55 \times 10^{10bc}$	1.12 × 10 <sup>11c</sup>	$1.22 \times 10^{9b}$	

Table 3 Changes in pH, syneresis percentage, and total plate count of yogurt samples during storage (4 °C)

<sup>a- c</sup>Mean values in the same row with different letters are significantly different (p < 0.05)

<sup>A, B</sup>Mean values in the same column with different letters are significantly different (p < 0.05)



#### Syneresis

As a commercial product, syneresis percentage is best kept at a minimum for yogurt to suit consumer preferences. Table 3 shows the effect of different MFDS forms addition to syneresis percentage in yogurt. The addition of MFDS forms gave a significant effect on the syneresis percentage of yogurt samples, although only yogurt with added MFDS powder experienced an increase in syneresis percentage by day 14. Syneresis percentages of yogurt with added MFDS powder  $(3.29 \pm 0.43 - 5.24 \pm 0.51\%)$ throughout storage are higher compared to other yogurt samples. Syneresis takes place due to the weakening of the yogurt gel structure thus losing the ability to entrap water (Sah et al. 2016). A higher syneresis percentage found in yogurt with added MFDS powder might be related to its TPC. Yogurt with MFDS powder reached the highest TPC out of all the samples  $(1.54 \pm 0.24 2.21\pm0.46$  mg/GAE g) and a high amount of phenolic compounds has been proven to affect yogurt syneresis by other studies. Increased rates of syneresis were detected in yogurts with added ingredients containing phenolic compounds such as blueberry juice, green tea powder, Ferulago angulata extract, and grape pomace (Marchiani et al. 2015; Jeong et al. 2018; Dimitrellou et al. 2020; Keshavarzi et al. 2020). Excess polyphenol concentrations could reduce the gel matrix that confines the yogurt serum by decreasing the volume of each protein-phenolic cage and preventing gel matrix formation (Jeong et al. 2018). Increasing the number of particle-particle junctions in the gel structure could lead to the shrinkage of the network and dismissing interstitial liquid (Dönmez et al. 2017).

#### Total plate count

Changes in the total plate count of yogurt with added MFDS are shown in Table 3. The addition of MFDS forms to yogurt did not give a significant difference in the total plate count in comparison to the control. The total plate count yogurt mixture increased after fermentation for all yogurt samples. However, the total plate count of yogurt with added MFDS did not increase significantly from day 0 to day 7. By day 14, the total plate count of all samples except the control experienced a decrease. Yogurt with added MFDS powder was not significantly affected during storage time even though it produced the same pattern of data as the other samples. The addition of MFDS powder to yogurt resulted in the lowest total plate count reached, from  $6.22 \times 10^9$  on day 0 to  $1.90 \times 10^7$  on day 14). Yogurt without any added MFDS experienced an increase in the total plate count by day 14, reaching  $1.96 \times 10^{11}$  from  $6.27 \times 10^9$  on day 0.

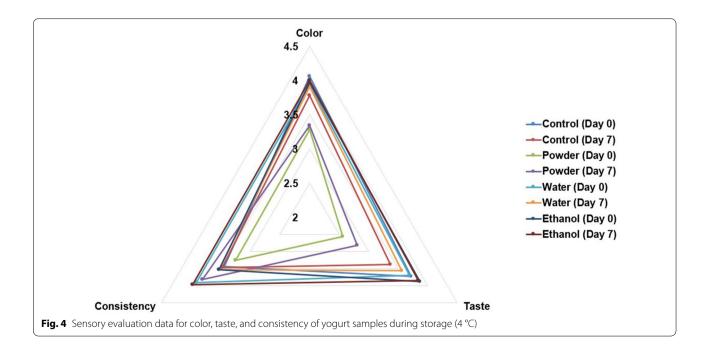
*Monascus* fermentation produces various compounds, among them are pigments and phenolics which have antibacterial characteristics. These compounds may have inhibited the growth of LAB during storage. Pigments are the major bioactive compounds found in *Monascus*fermented products along with monacolins (Zhu et al. 2019), with each group of pigments having different antibacterial mechanisms. Kim et al. (2006) found that amino acid derivatives of red *Monascus* pigments can suppress the growth of Gram-positive bacteria, which indicates that the red pigment derivatives from MFDS could have inhibited LAB growth. The hydrophobicity of bacteria cell surfaces increases when the pigments are adsorbed into the cell, which leads to cell aggregation into pellets. Pellet formation of cells results in the limited transfer of oxygen and nutrients into bacteria cells. Orange pigments have a good affinity with liposomes, resulting in an interaction with the phospholipid of bacteria cytoplasmic membranes. The interaction disrupts the bacteria membrane and causes cellular leakage. Orange pigments can also stimulate pellet formation in cells (Zhao et al. 2016). On the other hand, phenolics can disrupt cytoplasmic membrane and cause lysis, and interact with enzymes, substrates, and metal ions which prevents bacteria metabolism (Vaquero et al. 2007; Oulahal and Degraeve 2022; Melani et al. 2021).

MFDS water and ethanol extracts underwent further processing, starting with the extraction of compounds with various polarities using purely polar or nonpolar solvents and then being treated to sterilization. These caused the number of compounds in the extracts to be less than in the MFDS powder. This would explain why the addition of MFDS powder suppressed the growth of LAB in yogurt the most. Melani et al. (2021) found that the total plate count of *angkak*-supplemented kefir also decreased during storage time. Pyo and Song (2009), however, found that *Monascus*-fermented soybeans had an increase in viable LAB due to the free and essential amino acids produced from *Monascus* fermentation being a source of nutrients. According to the National Standardization Agency of Indonesia (BSN), the minimal amount of starter bacteria in yogurt is  $10^7 \log/g$  (BSN 2009) thus MFDS-supplemented yogurt is by the standard.

#### **Sensory evaluation**

According to Fig. 4, the acceptance scores of all yogurt samples were in the range of 2.56 (dislike-neutral) to 4.06 (like-most like). The addition of MFDS powder had a significant difference in the panelist acceptance score for color and taste, while the addition of any form of MFDS did not give a significant effect on the consistency acceptance score. Storage time did not give a significant effect on the panelist acceptance scores, which meant the quality of all yogurt samples remained stable after seven days of storage. Yogurt with added MFDS powder obtained the lowest acceptance scores for color (3.28-3.34) and taste (2.56-2.81) on days 0 and 7 respectively. The highest score for each parameter was reached by yogurt with added ethanol extract, with scores of 3.97-4.00 for color, 3.87-3.84 for taste, and 3.53-3.97 for consistency. The results show that the addition of MFDS, particularly the ethanol extract, to yogurt was well-liked by panelists.

Color analysis results from Table 2 show that yogurt with added MFDS powder produced a more redcolored yogurt in comparison to the other samples. Without the information about any flavor or added ingredients in yogurt, panelists could have an expectation of a white color regularly found in plain yogurt. A difference in the color of yogurt with added MFDS powder in comparison to other samples might affect

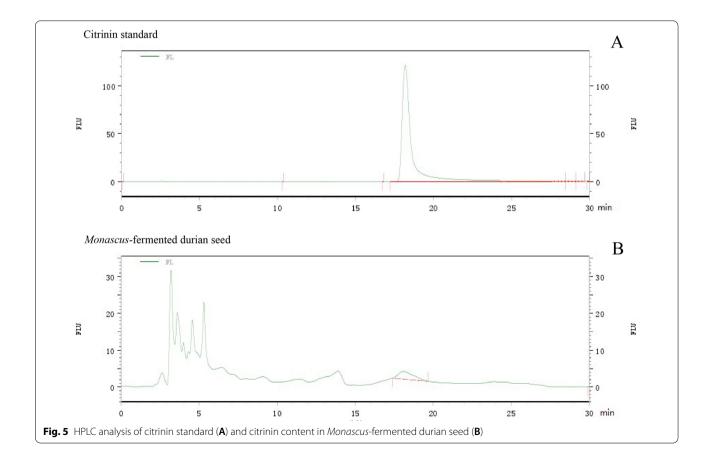


panelists' acceptance scores. MFDS powder was not incorporated evenly in the yogurt, which lead to the accumulation of powder at the bottom of the yogurt cup. This overall appearance might also affect the scores. Panelists have also commented that yogurt with added MFDS powder had a bitter taste. The bitterness might be due to the presence of phenolics in the yogurt (Li and Duan 2018). According to the total phenolic content data from Fig. 2, yogurt with added MFDS powder was found to have the highest TPC out of all the samples. Reginio et al. (2016) also reported that bitterness and astringency were detected by panelists from *Monascus* biopigment beverages.

#### Citrinin content of Monascus-fermented durian seed

A notable secondary metabolite from *Monascus purpureus* not yet discussed is citrinin. Citrinin is a mycotoxin produced from the same polyketide biosynthesis pathway as *Monascus* pigments and monacolin K (Agboyibor et al. 2018). Citrinin has cytotoxic, nephrotoxic, hepatotoxic, carcinogenic, and immunosuppression effects when ingested by humans and animals (Magro et al. 2016). It is essential to analyze the citrinin content of MFDS to ensure the safety of MFDS yogurt. HPLC analysis (Fig. 5) shows that the citrinin concentration of MFDS is 5.53 ppm.

Countries have different regulations for citrinin content in angkak. The citrinin limit in Taiwan is 2 ppm, 0.2 ppm in Japan, and 0.02 mg/kg body weight per day in Europe (Lee et al. 2010; Pattanagul et al. 2007; European Food Safety Authority 2012). The citrinin concentration of MFDS is higher than the limits mentioned, thus not recommended for high and frequent amounts of consumption. However, the MFDS extraction process with a 1:50 (w/v) ratio of MFDS and solvent produced MFDS yogurt with an estimated 0.1106 ppm of citrinin, which is below the limit in Taiwan and Japan. Citrinin has been stated to be unstable and thermolabile (Doughari 2015; Zhang et al. 2021). According to Trivedi et al. (1993a, b), citrinin heated at 80 °C for 60 min in watery conditions resulted in a 50% reduction but did not affect the citrinin cytotoxicity, while the heating temperatures of 90–110 °C increased the cytotoxicity. Citrinin H1, a highly toxic compound formed from citrinin degradation, was found after heating at 140 °C with the presence of water or at 100 °C for 30 min (Trivedi et al. 1993a, b). Shu and Lin (2002) found that citrinin concentration in angkak was dramatically decreased after boiling in water, while Hirota et al. (2002) stated that citrinin degrades on



heating above 80 °C under aqueous conditions. The processing heat of 80 °C during yogurt production would neither increase nor decrease the cytotoxicity of MFDS yogurt but might have slightly decreased citrinin content. Further research should be done regarding the amount of citrinin content in MFDS yogurt.

#### Conclusion

In this study, the addition of MFDS powder to yogurt produced the largest improvement in yogurt parameters, particularly its antioxidative properties, among the other forms of MFDS. Besides producing a more interesting appearance of yogurt from the red color MFDS imparts, TPC and antioxidant activity were also increased due to less loss of phenolic compounds in MFDS powder in comparison to other forms which undergo the extraction process. Due to higher phenolic content, yogurt with added MFDS powder also experienced the highest syneresis percentage and imparted a bitter taste, which was reflected in the lowest acceptance scores by panelists. MFDS powder had a high citrinin concentration. On the other hand, yogurt with added MFDS ethanol extract had the highest acceptance scores and the extracts had a lower citrinin concentration than the limits set. MFDS has the potential as a functional ingredient for food, especially yogurt.

#### Abbreviations

MFDS: *Monascus*-fermented durian seed; LAB: Lactic acid bacteria; GABA: γ-Aminobutyric acid; DPPH: Diphenyl-1-picrylhydrazyl; ACE: Angiotensinconverting enzyme; HPLC: High-performance liquid chromatography; LC–MS: Liquid chromatography–mass spectrometry; ABTS<sup>+</sup>: 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid); TPC: Total phenolic content.

#### Acknowledgements

Not applicable.

#### Author contributions

All authors read and approved the final manuscript.

#### Funding

Special thanks to Direktorat Riset dan Pengabdian Masyarakat, Deputi Bidang Penguatan Riset dan Pengembangan, Kementerian Pendidikan dan Kebudayaan, Riset dan Teknologi for the financial support for this research work with the contract number of 2600/WM01.5/N/2022.

#### Availability of data and materials

The datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

#### Declarations

#### **Competing interests**

The authors declare that they have no competing interests.

Received: 27 October 2022 Accepted: 2 December 2022 Published online: 14 December 2022

#### References

- Abd Razak DL, Abd Rashid NY, Jamaluddin A, Sharifudin SA, Long K (2015) Enhancement of phenolic acid content and antioxidant activity of rice bran fermented with *Rhizopus oligosporus* and *Monascus purpureus*. Biocatal 4(1):33–38. https://doi.org/10.1016/j.bcab.2014.11.003
- Abdel-Raheam HEF, Abdel-Mageed WS, Abd El-Rahman MAM (2019) Optimization of production of *Monascus ruber* pigments on broth medium and their application in flavored yogurts. Egypt J Food Sci 47(2):271–283. https://doi.org/10.21608/EJFS.2019.19654.1030
- Agboyibor C, Kong WB, Chen D, Zhang AM, Niu SQ (2018) *Monascus* pigments production, composition, bioactivity and its application: a review. Biocatal 16:433–447. https://doi.org/10.1016/j.bcab.2018.09.012
- Aloglu HS, Oner Z (2011) Determination of antioxidant activity of bioactive peptide fractions obtained from yogurt. J Dairy Sci 94(11):5305–5314. https://doi.org/10.3168/jds.2011-4285
- Amatayakul T, Sherkat F, Shah NP (2006) Syneresis in set yogurt as affected by EPS starter cultures and levels of solids. Int J Dairy Technol 59(3):221–261. https://doi.org/10.1111/j.1471-0307.2006.00264.x
- Aryana KJ, Olson DW (2017) A 100-year review: yogurt and other cultured dairy products. J Dairy Sci 12:9987–10013. https://doi.org/10.3168/jds. 2017-12981
- Baba AS, Najarian A, Shori AB, Lit KW, Keng GA (2014) Viability of lactic acid bacteria, antioxidant activity and in vitro inhibition of angiotensinl-converting enzyme of *Lycium barbarum* yogurt. Arab J Sci Eng 39(7):5355–5362. https://doi.org/10.1007/s13369-014-1127-2
- Bei Q, Liu Y, Wang L, Chen G, Wu Z (2017) Improving free, conjugated, and bound phenolic fractions in fermented oats (*Avena sativa* L.) with *Monascus anka* and their antioxidant activity. J Funct Foods 32:185– 194. https://doi.org/10.1016/j.jff.2017.02.028
- BSN (2009) SNI 2981:2009 Yogurt. Badan Standardisasi Nasional, Jakarta
- Chen C, Zhao S, Hao G, Yu H, Tian H, Zhao G (2017) Role of lactic acid bacteria on the yogurt flavor: a review. Int J Food Prop 20(1):316–330. https://doi.org/10.1080/10942912.2017.1295988
- Cheng J, Lee SK, Palaniyandi SA, Suh JW, Yang SH (2022) Effect of fermentation with *Monascus pilosus* on the antioxidant activities and phenolic acid contents of adzuki bean (*Vigna angularis*). J Coast Life Med 3(3):276–283. https://doi.org/10.12980/JCLM.3.2015JCLM-2015-0007
- Chouchouli V, Kalogeropoulos N, Konteles SJ, Karvela E, Makris DP, Karathanos VT (2013) Fortification of yoghurts with grape (*Vitis vinifera*) seed extracts. LWT Food Sci Technol 53(2):522–529. https://doi.org/10. 1016/j.lwt.2013.03.008
- Comunian TA, Chaves IE, Thomazini M, Moraes ICF, Ferro-Furtado R, Castro IA, Favaro-Trindade CS (2017) Development of functional yogurt containing free and encapsulated echium oil, phytosterol and sinapic acid. Food Chem 237:948–956. https://doi.org/10.1016/j.foodchem. 2017.06.071
- Darwish AZ, Darwish SM, Ismail MA (2017) Utilization of fermented yeast rice by the fungus *Monascus ruber* AUMC 4066 as food coloring agents. J Food Process Technol 8(1):1–6. https://doi.org/10.4172/2157-7110. 1000645
- Dimitrellou D, Solomakou N, Kokkinomagoulos E, Kandylis P (2020) Yogurts supplemented with juices from grapes and berries. Foods 9:1158. https://doi.org/10.3390/foods9091158
- Dönmez Ö, Mogol BA, Gökmen V (2017) Syneresis and rheological behaviors of set yogurt containing green tea and green coffee powders. J Dairy Sci 100(1):901–907. https://doi.org/10.3168/jds.2016-11262
- Doughari JH (2015) The occurrence, properties and significance of citrinin mycotoxin. J Plant Pathol Microbiol. https://doi.org/10.4172/2157-7471.1000321
- European Food Safety Authority (2012) Scientific opinion on the risks for public and animal health related to the presence of citrinin in food and feed. EFSA J 10(3):1–82. https://doi.org/10.2903/j.efsa.2012.2605
- Feng Y, Shao Y, Chen F (2012) *Monascus* pigments. Appl Microbiol Biotechnol 96(6):1421–1440. https://doi.org/10.1007/s00253-012-4504-3
- Gahruie HH, Eskandari MH, Mesbahi G, Hanifpour MA (2015) Scientific and technical aspects of yogurt fortification: a review. Food Sci Hum Wellness 4(1):1–8. https://doi.org/10.1016/j.fshw.2015.03.002
- Gjorgievski N, Yomovska J, Dimitrovska G, Makarijoski B, Shariati MA (2014) Determination of the antioxidant activity in yogurt. J Hyg Eng Des 8:88–92

- Hirota M, Menta AB, Yoneyama K, Kitabatake N (2002) A major decomposition product, citrinin H2, from citrinin on heating with moisture. Biosci Biotechnol Biochem 66(1):206–210. https://doi.org/10.1271/bbb.66.206
- Jeon CP, Lee JB, Choi CS, Kwon GS (2011) Physiological effect of yogurt with powder two stage fermented *Dioscorea batatas* Dence by *Monascus* sp. and *Lactobacillus* sp. Korean J Microbiol 47(2):151–157
- Jeong CH, Ryu H, Zhang T, Lee CH, Seo HG, Han SG (2018) Green tea powder supplementation enhances fermentation and antioxidant activity of set-type yogurt. Food Sci Biotechnol 27(5):1419–1427. https://doi. org/10.1007/s10068-018-0370-9
- Keshavarzi M, Sharifan A, Ardakani SAY (2020) Effect of the ethanolic extract and essential oil of Ferulago angulata (Schlecht.) Boiss. on protein, physicochemical, sensory, and microbial characteristics of probiotic yogurt during storage time. Food Sci Nutr 9(1):197–208. https://doi. org/10.1002/fsn3.1984
- Kim HS, Chae HS, Jeong SG, Ham JS, Im SK, Ahn CN, Lee JM (2005) Antioxidant activity of some yogurt starter cultures. Asian Aust J Anim Sci 18(2):255– 258. https://doi.org/10.5713/ajas.2005.255
- Kim C, Jung H, Kim YO, Shin CS (2006) Antimicrobial activities of amino acid derivatives of *Monascus* pigments. FEMS Microbiol Lett 264(1):117–124. https://doi.org/10.1111/j.1574-6968.2006.00451.x
- Lee BH, Pan TM (2012) Benefit of *Monascus*-fermented products for hypertension prevention: a review. Appl Microbiol Biotechnol 94(5):1151–1161. https://doi.org/10.1007/s00253-012-4076-2
- Lee CH, Lee CL, Pan TM (2010) A 90-D toxicity study of *Monascus*-fermented products including high citrinin level. J Food Sci 75(5):91–97. https://doi.org/10.1111/j.1750-3841.2010.01626.x
- Li SY, Duan CQ (2018) Astringency, bitterness and color changes in dry red wines befotr and during oak barrel aging: an updated phenolic perspective review. Crit Rev Food Sci Nutr 59(12):1840–1867. https://doi.org/10. 1080/10408398.2018.1431762
- Li FQ, Xu GR, Li YW, Chen Y (2003) Study on the production of citrinin by Monascus strains used in food industry. J Hygiene Res 32(6):602–605
- Liu D, Lv XX (2018) Effect of blueberry flower pulp on sensory, physicochemical properties, lactic acid bacteria, and antioxidant activity of set-type yogurt during refrigeration. J Food Process Preserv. https://doi.org/10. 1111/jfpp.13856
- Magro M, Moritz DE, Bonaiuto E, Baratella D, Terzo M, Jakubec P, Malina O, Čépe K, Aragao GMF, Zboril R, Vianello F (2016) Citrinin mycotoxin recognition and removal by naked magnetic nanoparticles. Food Chem 203:505–512. https://doi.org/10.1016/j.foodchem.2016.01.147
- Marchiani R, Bertolino M, Belviso S, Giordano M, Ghirardello D, Torri L, Piochi M, Zeppa G (2015) Yogurt enrichment with grape pomace: effect of grape cultivar on physicochemical, microbiological and sensory properties. J Food Qual 39(2):77–89. https://doi.org/10.1111/jfq.12181
- Melani D, Nurliyani N, Indratiningsih I (2021) Characteristics of goat milk kefir with addition of red yeast rice extract during storage. Bull Animal Sci 45(1):45–55. https://doi.org/10.21059/BULETINPETERNAK.V4511.12494
- Mirhosseini H, Amid BT, Cheong KW (2013) Effect of different drying methods on chemical and molecular structure of heteropolysaccharide-protein gum from durian seed. Food Hydrocoll 31:210–219. https://doi.org/10. 1016/j.foodhyd.2012.11.005
- Nugerahani I, Sutedja AM, Srianta I, Widharma RM, Marsono Y (2017) In vivo evaluation of *Monascus*-fermented durian seed for antidiabetic and antihypercholesterol agent. Food Res 1(3):83–88. https://doi.org/10.26656/fr. 2017.3.023
- Oulahal N, Degraeve P (2022) Phenolic-rich plant extracts with antimicrobial activity: An alternative to food preservatives and biocides? Frontiers Microbiol. https://doi.org/10.3389/fmicb.2021.753518
- Pattanagul P, Pinthong R, Phianmongkhol A (2007) Review of *angkak* production (*Monascus purpureus*). Chiang Mai J Sci 34(3):319–328
- Purnomo A, Yudiantoro YAW, Putro JN, Nugraha AT, Irawaty W, Ismadji S (2016) Subcritical water hydrolysis of durian seeds waste for bioethanol production. Int J Ind Chem 7:29–37. https://doi.org/10.1007/s40090-015-0059-3
- Pyo YH, Song SM (2009) Physicochemical and sensory characteristics of a medicinal soy yogurt containing health-benefit ingredients. J Agric Food Chem 57(1):170–175. https://doi.org/10.1021/jf8026952
- Ramli ANM, Sukri NAM, Azelee NIW, Bhuyar P (2021) Exploration of antibacterial and antioxidative activity of seed/peel extracts of Southeast Asian fruit Durian (*Durio zibethinus*) for effective shelf-life enhancement of preserved meat. J Food Process Preserv. https://doi.org/10.1111/jfpp.15662

- Reginio FJC, Hurtada WA, Dizon El (2016) Quality and acceptability of *Monascus* biopigment beverage. Int Food Res J 23(4):1492–1500
- Sah BNP, Vasiljevic T, McKechnie S, Donkor ON (2016) Physicochemical, textural and rheological properties of probiotic yogurt fortified with fibre-rich pineapple peel powder during refrigerated storage. LWT Food Sci Technol 65:978–986. https://doi.org/10.1016/j.lwt.2015.09.027
- Setyaningsih D, Apriyantono A, Sari MP (2010) Analisis sensori untuk industri pangan and agro. IPB Press, Bandung
- Shiby VK, Mishra HN (2013) Fermented milks and milk products as functional foods: a review. Crit Rev Food Sci Nutr 53(5):482–496. https://doi.org/10. 1080/10408398.2010.547398
- Shori AB (2013) Antioxidant activity and viability of lactic acid bacteria in soybean-yogurt made from cow and camel milk. J Taibah Univ Sci 7:202–208. https://doi.org/10.1016/j.jtusci.2013.06.003
- Shori AB, Baba AS (2013) Antioxidant activity and inhibition of key enzymes linked to type-2 diabetes and hypertension by *Azadirachta indica*-yogurt. J Saudi Chem Soc 17(3):295–301. https://doi.org/10.1016/j.jscs.2011.04. 006
- Shu PY, Lin CH (2002) Simple and sensitive determination of citrinin in *Monascus* by GC-selected ion monitoring mass spectrometry. Anal Sci 18(3):283–287
- Srianta I, Hendrawan B, Kusumawati N, Blanc PJ (2012) Study on durian seed as a new substrate for *angkak* production. Int Food Res J 19(3):941–945
- Srianta I, Kusumawati N, Nugerahani I, Artanti N, Xu GR (2013) In vitro α-glucosidase inhibitory activity of *Monascus*-fermented durian seed extracts. Int Food Res J 20(2):533–536
- Srianta I, Nugerahani I, Kusumawati N, Suryatanijaya E, Subianto C (2014) Therapeutic antioxidant activity of *Monascus*-fermented durian seed: a potential functional food ingredient. JJFNP 7(1):53–59. https://doi.org/10. 47556/J.JJFNPH.7.1.2014.5
- Srianta I, Zubaidah E, Estiasih T, Iuchi Y, Harijono YM (2017) Antioxidant activity of pigments derived from *Monascus purpureus*-fermented rice, corn, and sorghum. Int Food Res J 24(3):1186–1191
- Srianta I, Ristiarini S, Nugerahani I (2019) Pigments extraction from *Monascus*fermented durian seed. IOP Conf Ser Earth Environ Sci. https://doi.org/10. 1088/1755-1315/443/1/012008
- Srianta I, Kusdiyantini E, Zubaidah E, Ristiarini S, Nugerahani I, Alvin A, Iswanto N, Zhang BB (2021) Utilization of agro-industrial by-products in *Monascus* fermentation: a review. Bioresour Bioprocess. https://doi.org/10.1186/ s40643-021-00473-4
- Subianto C, Srianta I, Kusumawati N (2013) Effect of proportion of water and ethanol as solvent on antioxidant activity of durian seeds *angkak* with phosphomolybdenum and DPPH methods. J Food Technol Nutr 12(2):75–80. https://doi.org/10.33508/jtpg.v12i2.1487
- Suraiya S, Lee JM, Cho HJ, Jang WJ, Kim DG, Kim YO, Kong IS (2018) Monascus spp. fermented brown seaweeds extracts enhance bio-functional activities. Food Biosci 21:90–99. https://doi.org/10.1016/j.fbio.2017.12.005
- Szołtysik M, Kucharska AZ, Sokół-Łętowska A, Dąbrowska A, Bobak Ł, Chrzanowska J (2020) The effect of *Rosa spinosissima* fruits extract on lactic acid bacteria growth and other yoghurt parameters. Foods. https://doi. org/10.3390/2Ffoods9091167
- Tan H, Xing Z, Chen G, Tian X (2018) Evaluating antitumor and antioxidant activities of yellow *Monascus* pigments from *Monascus* ruber fermentation. Molecules. https://doi.org/10.3390/molecules23123242
- Trivedi AB, Doi E, Kitabatake N (1993a) Toxic compounds formed on prolonged heating of citrinin under watery conditions. J Food Sci 58(1):229–232. https://doi.org/10.1111/j.1365-2621.1993.tb03251.x
- Trivedi AB, Hirota M, Doi E, Kitabatake N (1993b) Formation of a new toxic compound, citrinin H1, from citrinin on mild heating in water. J Chem Soc Perkin Trans 1(18):2167–2171. https://doi.org/10.1039/P19930002167
- Vaquero MJR, Alberto MR, Nadra MCM (2007) Antibacterial effect of phenolic compounds from different wines. Food Control 18(2):93–101. https://doi.org/10.1016/j.foodcont.2005.08.010
- Virtanen T, Pihlanto A, Akkanen S, Korhonen H (2007) Development of antioxidant activity in milk whey during fermentation with lactic acid bacteria. J Appl Microbiol 102(1):106–115. https://doi.org/10.1111/j.1365-2672.2006. 03072.x
- Wang L, Luo Y, Wu Y, Wu Z (2018) Impact of fermentation degree on phenolic compositions and bioactivities during the fermentation of guava leaves with *Monascus anka* and *Bacillus* sp. J Funct Foods 41:183–190. https:// doi.org/10.1016/j.jff.2017.12.044

- Widagdha S, Nisa FC (2015) The effect of grape juice (*Vitis venifera* L.) and different fermentation period toward physico-chemical properties of yogurt. J Pangan Dan Agroindustri 3(1):248–258
- Zhang S, Liu L, Su Y, Li H, Sun Q, Liang X, Lu J (2011) Antioxidative activity of lactic acid bacteria in yogurt. Afr J Microbiol Res 5(29):5194–5201. https:// doi.org/10.5897/AJMR11.997
- Zhang H, Ahima J, Yang Q, Zhao L, Zhang X, Zhang X (2021) A review on citrinin: its occurrence, risk implications, analytical techniques, biosynthesis, physiochemical properties and control. Food Res Int. https://doi.org/10. 1016/j.foodres.2020.110075
- Zhao GP, Li YQ, Yang J, Cui KY (2016) Antibacterial characteristics of orange pigment extracted from *Monascus* pigments against *Escherichia coli*. Czech J Food Sci 34(3):197–203. https://doi.org/10.17221/430/2015-CJFS
- Zhihao W, Wen M, Yuhan Y, Gongjian F, Caie W, Tingting L, Chunyan X, Dongbei S (2022) Preparation of *Monascus*-fermented ginkgo seeds: optimization of fermentation parameters and evaluation of bioactivity. Food Sci Biotechnol 31:721–730. https://doi.org/10.1007/s10068-022-01078-z
- Zhu B, Qi F, Wu J, Yin G, Hua J, Zhang Q, Qin L (2019) Red yeast rice: a systematic review of the traditional uses, chemistry, pharmacology, and quality control of an important Chinese folk medicine. Front Pharmacol 10:1449–1476. https://doi.org/10.3389/fphar.2019.01449

#### **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

# Submit your manuscript to a SpringerOpen<sup>®</sup> journal and benefit from:

- Convenient online submission
- ► Rigorous peer review
- Open access: articles freely available online
- ► High visibility within the field
- ▶ Retaining the copyright to your article

Submit your next manuscript at > springeropen.com